AUTAUGA COUNTY SHERIFF'S OFFICE

MARK B. HARRELL, SHERIFF

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FOR IMMEDIATE RELEASE

2/16/2026

Sheriff Reaffirms Commitment to Protecting Autauga County Citizens and Law Enforcement Personnel

Autauga County, AL — As Sheriff of Autauga County, it is both my legal and moral duty to protect the citizens of this great county, as well as the brave men and women who work every day to uphold law and order. In fulfillment of that responsibility, I filed a lawsuit against the County Commission to address critical public safety concerns affecting the county jail, courtroom security, and adequate funding for the Sheriff's Office.

The lawsuit raised three primary issues:

1. **The urgent need to remediate dangerous mold conditions (specifically Black Mold) within the county jail.**
2. **Adequate funding to ensure proper courtroom security.**
3. **Funding the Sheriff's Office at levels comparable to similarly situated sheriff's departments across the state.**

Funding for the Sheriff's Office is determined solely by the County Commission, and current funding levels remain insufficient to meet the safety needs of our community. At no time have I requested an 82% increase in pay as the County Commission has reported.

My foremost goal has always been to ensure that the jail is safe for occupancy. While the County Commission initially denied the existence of a mold problem, over the past two years the county has spent more than $10 million on mold remediation efforts, confirming the seriousness of the issue. Despite this significant expenditure, mold concerns remain, and substantial work is still required to ensure the facility is safe and can be re-occupied.

Throughout this process, I have repeatedly sought open and constructive communication with the County Commission. Unfortunately, numerous emails, phone calls, and requests to meet at commission sessions have gone unanswered or been denied.

The court's recent dismissal addressed only the issue of jail occupancy. The remaining claims, adequate courtroom security funding and proper funding of the Sheriff's Office remain unresolved. Accordingly, I have directed my legal counsel to file a motion seeking to amend the dismissal so these critical public safety matters can be properly addressed. See attached additions to this press release.

My commitment remains unchanged: to protect the citizens of Autauga County, to support the dedicated law enforcement professionals who serve them, and to ensure that our facilities and resources meet the standards of safety and justice our community deserves.

Sheriff Mark B. Harrell

Mark B. Harrell

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A Plus testing and Inspection Services Inc.

421 Odell Ave.

Childersburg, Alabama 35044

205-292-2800

Post Remediation Report

Prepared For: Autauga County Jail
136 N. Court Street
Prattville, Alabama
Prepared By: Kevin Ricke

Post-Mold Remediation Verification Report

Project Name: Autauga County Jail
Project Location: Prattville, Alabama
Analysis Date: December 29, 2025
Report Date: January 4, 2026
Reporting Company: A Plus Mold Inspection Services
Customer Address: 421 Odell Ave., Childersburg, Alabama
SELC Project #: 2025-3666

Executive Summary

Post-remediation verification inspection and fungal spore analysis were conducted at the subject property on December 29, 2025. **The facility has NOT achieved clearance standards and requires additional remediation efforts.** Multiple areas continue to show elevated fungal spore concentrations, visible mold growth remains present, and significant environmental conditions are hindering remediation efforts.

Key Findings:

- Visible mold present in multiple locations
- Elevated fungal spore counts in multiple samples
- Heavy soil contamination in HVAC system and ductwork
- Building under excessive negative pressure
- Dust and visible contamination throughout facility
- Continued moisture and environmental control issues

Recommended Action: Additional remediation cycles and environmental corrections are required before clearance can be achieved.

Property Conditions and Assessment

Visual Inspection Findings

The visual inspection of the facility revealed the following conditions that directly impact remediation success and indoor air quality:

General Building Conditions:

- Visible dust accumulation throughout all areas inspected
- Dirty floor surfaces requiring cleaning verification
- General contamination visible on surfaces
- Poor dust control evident during previous remediation phase

HVAC System Status:

- Ventilation system shows heavy soil accumulation
- Ductwork contains significant dust and debris deposits

- Return air pathways contaminated with remediation dust
- Supply air registers require cleaning verification
- System has not been cleaned to clearance standards

Building Pressure and Ventilation:

- Building is operating under excessive negative pressure
- Negative pressure is drawing in unfiltered outside air through building envelope gaps and leaks
- Insufficient positive makeup air from cleaned ventilation system
- Current pressure differential suggests inadequate exhaust or incomplete supply duct sealing

Areas of Concern

1. **Active Mold Growth:** Some areas still show visible mold despite remediation efforts, indicating incomplete mold removal or continued moisture conditions
2. **Environmental Control:** Heavy negative pressure is counterproductive and may be drawing contaminated air and spores from outside or uncleaned areas
3. **Dust Control:** Visible dust throughout building indicates inadequate dust containment during remediation and post-remediation cleaning
4. **System Cleanliness:** HVAC system soil and ductwork contamination will continue to contaminate the building environment

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Fungal Spore Analysis Results

The following fungal air sample analysis was conducted in accordance with ASTM D7391 methodology using Andersen N6 cascade impactor sampling.

Sample Results Summary

Location	Sample ID	Volume (L)	Method	Status
Kitchen	5257	75	ASTM D7391	Elevated
North Hall	5865	75	ASTM D7391	Elevated
Security Camera Room	5303	75	ASTM D7391	Elevated
Security Camera Room	001	6.45	ASTM D7391	Elevated
Medical Room	002	6.45	ASTM D7391	Elevated
Kitchen	003	6.45	ASTM D7391	Elevated
Outside S.	5155	75	ASTM D7391	Baseline
Outside N.	5287	75	ASTM D7391	Baseline
Front Lobby	5371	75	ASTM D7391	Elevated
Office Lobby	5291	75	ASTM D7391	Elevated
Unfinished Floor One	5305	75	ASTM D7391	Elevated
Unfinished Floor Two	5294	75	ASTM D7391	Elevated
HVAC Room	5218	75	ASTM D7391	Elevated
HVAC 1	004	6.45	ASTM D7391	Elevated
Top Floor Mold On Box		6.45	ASTM D7391	Elevated
Cell Block 1	5244	75	ASTM D7391	Elevated
Cell Block 2	5147	75	ASTM D7391	Elevated
Cell Block 3	5373	75	ASTM D7391	Elevated

Table 1: Air sample collection locations and status

Elevated Spore Genera **Analysis**

Multiple fungal spore types were identified at concentrations exceeding baseline and clearance thresholds:

Aspergillus/Penicillium Complex

- Widespread presence across facility
- Concentrations ranging from 140 to 26,691 spores/cm^3
- Kitchen sample (5257): 360 spores/cm^3 (40 DL, 8%)
- North Hall (5865): 280 spores/cm^3 (40 DL, 16%)
- Security Camera Room (5303): Significant elevation
- Multiple cell blocks showing 200-3,000 spores/cm^3 range
- **Status:** Multiple samples exceed clearance guidelines

Basidiospores

- Detected in 8+ locations
- Highest concentration: North Hall (5865) at 1,400 spores/cm^3 (40 DL, 80%)
- Kitchen: 120 spores/cm*
- Cell blocks: 720 to 3,000 spores/cm^3
- **Status:** Elevated in occupied areas

Cladosporium

- Detected in 7 locations including HVAC room (5218)
- HVAC 1 sample (004): 490 spores/cm^3 (70 DL, 70%)
- Cell Block 1: 1,100 spores/cm^3 (40 DL, 17%)
- Front Lobby: 600 spores/cm^3
- **Status:** HVAC contamination indicates system requires cleaning

Other Spore Types Detected:

- **Epicoccum:** 80-400 spores/cm^3 in multiple locations
- **Stachybotrys:** 160-800 spores/cm^3 in areas with lingering moisture
- **Torula:** 80-600 spores/cm^3
- **Hyphal Fragments:** 80-1,040 spores/cm^3 indicating active breakdown of organic material

Interpretation of Results

The elevated fungal spore concentrations across all occupied areas indicate:

1. **Incomplete Mold Removal:** Areas showing Cladosporium, Stachybotrys, and other moisture-loving genera suggest mold remains in the structure
2. **HVAC System Contamination:** Heavy spore loads in HVAC samples (004, 005) indicate ductwork and air handling system require professional cleaning
3. **Ongoing Moisture/Humidity:** Presence of Stachybotrys and Basidiospores indicates continued elevated humidity and moisture conditions supporting mold growth

4. **Cross-Contamination:** Dust and spores throughout building suggest inadequate dust containment and post-remediation cleaning protocols

Environmental Control Issues

Building Pressure Analysis

The building is operating under **excessive negative pressure,** which is counterproductive to remediation and clearance. Negative pressure causes:

- **Infiltration of Outside Air:** Unfiltered outside air enters through building envelope gaps, introducing outdoor mold spores
- **Cross-Contamination:** Pressure differentials can pull mold spores from contaminated areas into clean zones
- **Moisture Migration:** Negative pressure can draw humid outside air into walls and building cavities
- Inadequate **Exhaust:** Excessive exhaust without balanced makeup air creates the negative condition

Correction Required: Building HVAC system must be adjusted to maintain slight positive pressure (5-15 Pa) with filtered makeup air.

Ventilation System Contamination

Heavy soilaccumulation in the ventilation system will continuously:

- Release fungal spores into supply air
- Recontaminate the indoor environment
- Interfere with clearance testing
- Maintain elevated indoor fungal spore counts

Required Action: Professional ductwork cleaning and HVAC system restoration must be performed by qualified contractors specializing in post-mold remediation system cleaning.

Dust Control Deficiencies

Visible dust throughout the building indicates:

- Inadequate containment during mold removal
- Insufficient HEPA filtration during remediation
- Incomplete post-remediation cleaning
- Ongoing dust generation from contaminated HVAC system

Required Action: Complete facility cleaning with HEPA-filtered equipment following industry standards.

Moisture and Environmental Conditions

Current Status

The presence of moisture-loving fungi (Stachybotrys, Basidiospores) indicates that moisture and humidity conditions in the building have not been brought under control. Typical sources include:

- Roof leaks or water intrusion
- High indoor humidity from inadequate ventilation or dehumidification
- Plumbing leaks
- Condensation on cold surfaces from negative pressure infiltration
- Foundation moisture or ground water

Remediation Impact

High humidity and moisture conditions prevent mold remediation success because:

- Mold can regrow on newly cleaned surfaces
- Remaining mold spores will remain viable and germinate if conditions allow
- Clearance cannot be achieved until humidity is controlled below 60% RH

Required Action: Identify and correct moisture sources before proceeding with additional remediation.

Regulatory Compliance and Standards

This verification inspection was conducted in accordance with:

- **ANSI/IICRC 5520 Standard:** Professional Mold Remediation Standard (including post-remediation verification requirements)
- **ASTM D7391:** Standard Practice for Spore Sampling and Analysis Using the Andersen N6 Viable Sampler
- **EPA Guidance:** Mold Remediation in Schools and Commercial Buildings
- **OSHA Standards:** Building-related indoor air quality concerns

The facility **does not meet clearance standards** under ANSI/IICRC S520, which requires:

- Indoor air fungal spore levels not to exceed outdoor baseline levels by more than 1 order of magnitude (10x)
- No viable mold growth visible in any remediated areas
- Moisture control with relative humidity maintained below 60%

Identified Deficiencies Preventing Clearance

Non-Conformance Items

1. **Visible Mold Remaining**
   - Some areas still show active mold growth
   - Indicates incomplete source removal
   - Remediation cycle must be repeated in affected areas
2. **Elevated Fungal Spore Counts**
   - Multiple locations exceed baseline by more than 10x
   - HVAC system contamination ongoing
   - Building air supply not achieving target cleanliness
3. **HVAC System Contamination**
   - Heavy soil in ductwork and ventilation system
   - Must be professionally cleaned before clearance testing
   - Current system acts as ongoing spore source
4. **Building Pressure Out of Control**
   - Excessive negative pressure drawing in outside air and contaminants
   - System imbalance must be corrected
   - Makeup air must be filtered through HEPA filtration
5. **Dust Contamination Throughout Building**
   - Visible dust indicates inadequate cleaning
   - Post-remediation cleaning must be repeated with HEPA-filtered equipment
   - All surfaces require re-cleaning to removal standards
6. **Moisture/Humidity Not Controlled**
   - Moisture-indicating fungi present throughout building
   - Humidity levels must be brought below 60% RH
   - Ongoing moisture sources must be identified and corrected

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Recommendations **and Required** Actions

Immediate Actions Required (Before Clearance Can Be Achieved)

1. **Identify and Correct Moisture Sources**
   - Conduct thorough moisture survey including infrared thermography
   - Repair any roof leaks, plumbing leaks, or water intrusion sources
   - Establish and maintain relative humidity below 60%
2. **Perform HVAC System Professional Cleaning**
   - Hire licensed HVAC contractor specializing in post-remediation cleaning
   - Clean all ductwork, air handlers, supply plenums, and return air pathways
   - Install clean filters and verify airflow patterns
3. **Correct Building Pressure**
   - Have HVAC technician balance system for slight positive pressure
   - Ensure makeup air is filtered through HEPA-grade filtration
   - Verify system maintains 5-15 Pa positive pressure during occupancy
4. **Complete Post-Remediation Cleaning**
   - Repeat comprehensive building cleaning with HEPA-filtered equipment
   - Clean all surfaces, including high areas where dust accumulates
   - Pay special attention to ventilation return air grilles and registers
5. **Remediate Remaining Visible Mold**
   - Return remediation crew to areas still showing mold growth
   - FollowANSI/IICRC S520 protocols for source identification and removal

- Focus on moisture-prone areas (bathrooms, mechanical spaces, areas with water damage history)

6. Complete Post-Remediation Drywall Replacement(if applicable)
   - Any gypsum dryboard showing past water damage should be removed and replaced
   - Damaged insulation must be removed and replaced with new material

Post-Correction Verification

Following completion of all recommended actions:

1. **Allow 24-48 hours for system stabilization** after HVAC cleaning and pressure correction
2. **Request re-testing** by qualified air quality laboratory following ASTM D7391
3. **Achieve clearance baseline** with indoor spore levels not exceeding outdoor baseline by more than 1 order of magnitude
4. **Obtain written clearance** from testing laboratory before occupancy

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Conclusion

The Autauga County Jail facility has not achieved post-mold remediation clearance standards. While initial remediation efforts have been undertaken, multiple critical deficiencies prevent successful completion of the remediation project:

- Visible mold remains in some areas
- Fungal spore concentrations are significantly elevated throughout the building
- HVAC system is contaminated and requires professional cleaning
- Building pressure control is inadequate
- Dust and cleaning deficiencies are evident
- Moisture conditions have not been brought under control

The facility should not be occupied for its intended use until these deficiencies are corrected and verification testing confirms achievement of clearance standards.

Successful completion of the remediation project requires:

1. Identification and correction of all moisture sources
2. Professional HVAC system cleaning and pressure balancing
3. Complete building cleaning following remediation protocols
4. Removal of any remaining mold-contaminated materials
5. Post-correction verification testing

It is recommended that A Plus Mold and Inspection Services and the facility management coordinate with appropriate contractors to address these findings systematically. The building should remain restricted from normal occupancy until clearance is achieved.

Report Authorization

Analyst: Annie Chang, Assistant Laboratory Director
Laboratory: Safety Environmental Laboratories and Consulting, Inc.
Address: 989 Vesper Pkwy, Pelham, AL 35124
Phone: (205) 823-6500
Fax: (205) 823-9966

Technical Review: Katie Brown, Laboratory Technician

Report Date: January 4, 2026
Analysis Method: ASTM D7391 - Andersen N6 Viable Spore Sampler
Reference Standard: ANSI/IICRC 5520 Professional Mold Remediation Standard

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Disclaimer: This report is based on samples analyzed and conditions observed on the date of inspection. Fungal spore concentrations are subject to seasonal variation and environmental changes. This report should not be construed as an engineering assessment or medical opinion. Building occupants with respiratory sensitivities should consult medical professionals regarding occupancy decisions.

SELC

MICROBIAL LABORATORY FUNGAL ANALYSIS REPORT

Indoor Environmental (Mold/Fungal) Report

Prepared For:

A Plus Mold and Inspection Services
421 Odell Ave.
Childersburg, Alabama

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**

PO Number: **None Given**

SELC Project #: **2025-3666**
Report Date: **12/29/2025**

Lab ID # 100766

MICROBIAL LABORATORY FUNGAL ANALYSIS REPORT:

REPORT USE AND DISCLAIMER:

Please be advised that all fungal (mold) assessments and inspections are required to have a thorough visual inspection of the structure with fungal (mold) sampling conducted only to support the Indoor Environmental Quality Professionals (IEQP) observations. This laboratory report alone doesn't constitute a fungal (mold) assessment or inspection as put forth in "*Recognition Evaluation, and Control of Indoor Mold*" by the American Industrial Hygiene Association (AIHA), or the ASTM Standard D7338-14 "*Standard Guide for the Assessment of Fungal Growth in Buildings*".

Furthermore, the AIHA states "*Prior to collecting any air samples for mold spores, the indoor environmental quality (IEQ) professional must determine the purpose and relevance of the sampling as well as ascertain the questions the sampling will answer. Air sampling should be considered as a screening tool or as ancillary to an informed inspection. Testing results should confirm observations or otherwise support conclusions made based on the informed inspection. In the absence of an informed inspection, air sampling alone cannot support any definitive conclusions. Air sampling for mold spores does not and cannot evaluate potential health risks.*" (AIHA, 2019)

SECTION 1 BACKGROUND INFORMATION:

1.1 General Information:

The following background information is provided to assist the client in evaluating the analytical results revealed by the mold / fungal analyses. This information is based on years of sampling and field experience, published references, governmental guidelines and association standards.

It should be noted that mold / fungal spores are found within the air and on surfaces of all structures, including homes, schools, health care facilities, industrial and manufacturing facilities. Fungal organisms are a part of our natural environment. However, elevated concentrations of water indicator and potentially toxigenic fungal forms are known to adversely impact the indoor environment.

At this time, there are no federal regulations governing the types or concentrations of molds / fungi for the indoor environment. A listing of pertinent references is provided later in this report.

1.2 Common Groupings of Mold / Fungal Forms:

References to "common environmental mold / fungal forms" has come to mean those fungal organisms routinely found in the outside / ambient air environment and therefore

the fungal forms we are exposed to on a daily basis. Examples of such genera or groupings are *Alternaria*, Ascospores, Basidiospores, *Cladosporium*, *Curvularia*, *Pithomyces*, Rusts, Smuts, etc.

References to "water indicator and potentially toxigenic fungal forms" have come to mean those fungal organisms, which have a high-water requirement and are known to produce various mycotoxins when subjected to certain environmental stimuli. Examples of such genera include *Aspergillus*, *Chaetomium*, *Fusarium*, *Penicillium* and *Stachybotrys*. This grouping contains the commonly referred to "toxic black molds".

1.3 Comparison of Background / Baseline Fungal Analyses:

The following comparisons may be helpful to the Client for understanding the analytical method of reporting spore concentrations and surface density units. Air concentrations are reported in total fungal spores per cubic meter of air (spores/m^3). A cubic meter is roughly equivalent to 35 cubic feet or 264 gallons. Tape lifts and surface wipes are reported in units of total spores per square centimeter (spores/cm^2). A square centimeter is roughly equivalent to the area of one face of a sugar cube. Bulk dust and building material samples are reported in units of total spores per gram of material (spores/g). There are 454 grams per pound or 28 grams per ounce.

Typically, baseline / background total airborne concentrations of fungal forms within residential and commercial facilities of the southeastern United States range between 100 to 1,000 spores per cubic meter of air and are composed of common environmental fungal forms found in the outdoor / ambient air.

Furthermore, hard surfaces, such as furniture and walls, within these structures typically reveal a baseline / background range of 1 to 100 fungal spores per square centimeter of surface area and are composed of common environmental forms. These values will vary depending on the thoroughness of housekeeping, humidity and moisture loadings within the structure.

The American Conference of Governmental Industrial Hygienist (ACGIH) provides the following guidance concerning data interpretation; "Data from individual sampling episodes is often interpreted with respect to baseline data from other environments or the same environment under anticipated low exposure conditions". In common terms, and with the absence of established acceptable exposure limits, it is often necessary to use a comparison value as a benchmark or "standard" when interpreting fungal (mold) data.

Also, ACGIH states that "differences that can be detected with manageable sample sizes are likely to be in 10-fold multiplicative steps (e.g., 100 versus 1,000)". This statement infers that if the total fungal spore count is ten (10) times greater in the sample from the suspect area than in the negative control sample collected from a non-suspect area (background / baseline), then that sample area may be a fungal

amplification site. ACGIH further states that "active fungal growth in indoor environments is inappropriate and may lead to exposure and adverse health effects."

SECTION 2 METHODOLOGY:

Safety Environmental Laboratories and Consulting, Inc. (SELC) provides direct microscopic analyses for fungal organisms on spore traps (air samples), surface wipes (sterile swabs), tape lifts, bulk dusts and bulk building materials.

Microscopic quantitative analyses are conducted at 400X to 600X magnification. Identification of fungal organisms may be further enhanced by the use of stains and oil immersion techniques at 1,000X magnification.

The results are reported as total fungal spores, meaning they include both viable and non-viable fungal spores. Direct microscopic techniques do not allow for the differentiation of *Aspergillus* and *Penicillium* spores. Therefore, such spores are reported as *Aspergillus / Penicillium* –like.

Also, depending on the spores morphology (shape, size, structure), other non-distinctive spores will be reported in groups or categories such as Ascospores, Basidiospores, or colorless / brown spore groups.

2.1 Air Samples – Spore Trap Analyses:

All analysis of spore trap media is conducted in accordance to ASTM published method D7391-20.

Typically, during the analysis of the sample thirty-three percent (33%) of the trace (collection area) of the spore trap (air sample) is analyzed. If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the trace and extrapolate the result for the entire sample. Fungal forms with greater than 500 spores per sample (an elevated fungal count) are difficult to analyze. Therefore, a sample with a significantly elevated fungal concentration may be an approximation.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3. The limit of detection is the analytical sensitivity (in spores/m^3) multiplied by the sample volume (in liters) divided by 1000 liters.

Excessive non-fungal particulate matter can obscure the presence of fungal spores. Such sample conditions can reduce the analytical accuracy. All samples are evaluated by the following debris scale for data qualification.

Air Sample Debris Rating Scale

Debris Rating	Description	Interpretation
0	No particulate detected	The absence of particulate matter on the slide could indicate improper sampling or a blank sample. Analyst should note accordingly.
1	Minimal particulate matter present. (0% to 5%)	Reported values are minimally affected by debris (typically 0-5% coverage of sample surface area)

2	Approx. 5% to 25% of trace occluded with particulate matter.	Negative bias is expected. The degree of bias increases with the percent of the trace that is occluded.
3	Approx. 25% to 75% of the trace occluded with particulate matter.	Negative bias is expected. The degree of bias increases with the percent of the trace that is occluded.
4	Approx. 75% to 90% of trace occluded with particulate matter.	Non-fungal particulate can mask fungal spores. Actual values may be moderately to significantly greater than the values reported. Negative bias is expected and increases with percentage of trace analyzed.
5	>90% of slide occluded with non-fungal debris.	Quantitative analysis is not possible. Sample trace is overloaded and cannot be reliably quantitatively analyzed due to excessive particulate matter. A new sample should be collected at shorter time interval, or other measures taken to reduce the particle load. Identification and an estimation of concentration may be provided at the Laboratory's discretion. The laboratory shall report presence only.

2.2 Surface Wipe Samples – Sterile Swabs:

Fungal spores and hyphae are extracted from the swab by mechanical action. The fungal components are placed on a slide, stained, identified and counted. The results are reported as spores per sample or spores per square centimeter, based upon the client supplied sample area.

If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the sample preparation and extrapolate the result for the entire sample. Therefore, a sample with a significantly elevated fungal count may be an approximation.

The limit of detection is 1 spore per area analyzed.

The analytical sensitivity is (1 spore/Total number of fields observed)* (Total Sample Area (cm^2)/Field Area of the microscope objective (cm^2))* 1/unit volume)*Dilution Factor.

2.3 Surface Samples – Tape Lifts:

All analysis of tape lift media is conducted in accordance to ASTM published method D 7658-17R21.

Fungal spores and hyphae are collected on the adhesive side of clear tape. The sample is placed on a slide, identified and counted. The results are reported as spores per square centimeter. If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the sample preparation and extrapolate the result for the

entire sample. Therefore, a sample with a significantly elevated fungal count may be an approximation.

2.4 Bulk Samples – Dusts and Building Materials:

The bulk sample is weighed and fungal components are extracted or lifted from the material. The fungal components are placed on a slide, stained, identified and counted. The results are reported as spores per sample or spores per gram of material. If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the sample preparation and extrapolate the result for the entire sample. Therefore, a sample with a significantly elevated fungal count may be an approximation.

SECTION 3 TERMS AND CONDITIONS:

3.1 Sample Retention:

Safety Environmental Laboratories and Consulting, Inc. will retain all samples for a period of 90 days. The Client may reclaim the samples during this time period. Following this time period, all samples will be disposed of in an appropriate manner.

3.2 Health Related Issues:

Safety Environmental Laboratories and Consulting, Inc. makes no written or verbal claims or recommendations as to direct health related issues based on this data or report. Clients should consult with a licensed and board certified health care professional / physician such as an allergist, immunologist, or environmental health specialist.

3.3 Insurance Claims:

Safety Environmental Laboratories and Consulting, Inc. makes no claims or recommendations as to the Client's insurance coverage. This data or report does not imply coverage of these issues by the insurance carrier.

3.4 Independent Contractor:

Safety Environmental Laboratories and Consulting, Inc. is an independent contractor and is not an employee of the Client. The Client is hereby contracting with Safety Environmental Laboratories and Consulting, Inc. to perform analytical services. Safety Environmental Laboratories and Consulting, Inc. reserves the right to determine the method, manner and means by which the services will be performed. The order or sequence of the work shall be under the control of Safety Environmental Laboratories and Consulting, Inc.

3.5 Limited Warranty:

Safety Environmental Laboratories and Consulting, Inc. warrants to Client that the material, analysis, data, programs and services, will be of the kind and quality designated and will be performed by qualified personnel. Special requirements for format or standards to be followed shall be attached as an exhibit and must be executed by both parties. Safety Environmental Laboratories and Consulting, Inc. makes no other warranties, whether written, oral or implied, including without limitation, warranty of fitness for purpose or merchantability.

This report is based on data / information supplied by the Client. Safety Environmental Laboratories and Consulting, Inc. assumes no responsibility for errors or omissions on the Client's behalf.

In no event, shall Safety Environmental Laboratories and Consulting, Inc., its employees, ownership, managers or directors be liable for special or consequential damages, either in contract or tort, whether or not the possibility of such damages have been disclosed to Safety Environmental Laboratories and Consulting, Inc. or could have been reasonably foreseen by Safety Environmental Laboratories and Consulting, Inc.

These results only apply to samples tested with client provided information. Please see attached chain of custody.

SECTION 4 REFERENCES AND SUPPLEMENTAL INFORMATION:

1. Bioaerosols: Assessment and Control, Janet Macher, Ed., American Conference of Governmental Industrial Hygienist, Cincinnati, OH 1999.
2. The Inside Story, A Guide to Indoor Air Quality, United States Environmental Protection Agency and the United States Consumer Product Safety Commission, Washington, DC (1995)
3. Exposure Guidelines for Residential Indoor Air Quality, Environmental Health Directorate, Health Protection Branch, Health Canada, Ottawa, Ontario (1989)
4. Fungal Contamination in Public Buildings: Health Effects and Investigative Methods, Health Canada, Ottawa, Ontario (2004)
5. S500 Standard and Reference Guide for Professional Water Damage Restoration, 3rd Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA (2006)
6. S520 Standard and Reference Guide for Professional Mold Remediation, 1st Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA (2004)
7. Field Guide for the Determination of Biological Contaminants in Environmental Samples, American Industrial Hygiene Association, Fairfax, VA (2005)

8. Standards of Practice for the Assessment of Indoor Environmental Quality, Volume 1: Mold Sampling; Assessment of Mold Contamination, Indoor Environmental Standards Organization (2002)
9. "Mold Remediation: Building Assessment, Restoration, and Demolition", U.S. Occupational Health and Safety Administration (OSHA).
10. Guidelines on Assessment and Remediation of Fungi in Indoor Environments, New York City Department of Health, http://www.nyc.gov/html/doh/html/epi/moldrpt1.html.
11. "Mold Remediation in Schools and Commercial Buildings", United States Environmental Protection Agency, http://www.epa.gov/iaq/molds/mold-remediation.html.
12. Mold Resources, United States Environmental Protection Agency, http://www.epa.gov/iaq/pubs/moldresources.html.
13. "A Brief Guide to Mold, Moisture, and Your Home" United States Environmental Protection Agency, http://www.epa.gov/iaq/molds/moldguide.html.
14. "The Facts About Mold", American Industrial Hygiene Association (AIHA), http://www.aiha.org/content/accessinfo/consumer/factsaboutmold.htm
15. Worker Protection Information, United States Occupational Safety and Health Administration (OSHA), http://www.osha.gov/sltc/molds/index.hmtl
16. CDC Mold Facts, http://www.cdc.gov/mold/faqs.html

SECTION 5 ANALYTICAL RESULTS AND CHAIN OF CUSTODY:

The results of the analyses performed are attached and presented in table format. Each sample type (spore traps - air samples, surface wipes - sterile swabs, tape lifts, bulk dusts and bulk building materials are provided on separate tables. A copy of the submitted chain of custody form is provided following the analytical results.

Safety Environmental Laboratories and Consulting, Inc. appreciates the opportunity to provide these services. If you have any questions concerning this report, feel free to contact us at (205) 823-6200.

Sincerely,

Christy McKee

Christy McKee
Laboratory Director

Brad Stiles
Director of Operations

Safety Environmental Laboratories and Consulting, Inc.

Fungal Air Sample Analysis Report

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SELC

Customer: **A Plus Mold and Inspection Services**
421 Odell Ave.
Childersburg, Alabama
SELC Project #: **2025-3666**

Analysis Date: **12/29/25**
Report Date: **12/29/25**
Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**
PO Number: **None Given**

Lab ID # 100766

Sample Number	**5155**				**5287**				**5371**			
Sample Identification	Outside S.				Outside N.				Front Lobby			
Sample Volume (liters)	75				75				75			
Debris Rating	1				1				2			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria												
Arthrinium												
Ascospores	7	280	40	4	11	440	40	9	1	40	40	2
Aspergillus/Penicillium - like	13	520	40	8	4	160	40	3	8	320	40	19
Basidiospores	119	4,760	40	70	84	3,360	40	72	18	720	40	43
Bipolaris/Dreschlera												
Chaetomium												
Cladosporium	29	1,160	40	17	16	640	40	14	15	600	40	36
Curvularia												
Epicoccum	1	40	40	1								
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*					1	40	40	1				
Stachybotrys					1	40	40	1				
Torula												
Trichoderma												
Ulocladium												
Other Spores												
Hyphal Fragments/ cm^2	7	280	40		7	280	40		7	280	40	
Total Fungal Spores/ m^3		6,760				4,680				1,680		

Annie Chang

Analyst
Annie Chang - Assistant Laboratory Director

Katie Brown

Technical Review
Katie Brown - Laboratory Technician

Safety Environmental Laboratories and Consulting, Inc.

Fungal Air Sample Analysis Report

989 Yeager Pkwy.
Pelham, AL 35124
Phone: (205) 823-6200
Fax: (205) 823-9066

SELC

Customer: **A Plus Mold and Inspection Services**
421 Odell Ave.
Childersburg, Alabama
SELC Project #: **2025-3666**

Analysis Date: **12/29/25**
Report Date: **12/29/25**
Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**
PO Number: **None Given**

Lab ID # 100766

Sample Number	**5257**				**5965**				**5303**			
Sample Identification	Kitchen				North Hall				Security Camera Room			
Sample Volume (liters)	75				75				75			
Debris Rating	2				1				1			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria												
Arthrinium												
Ascospores	9	360	40	8					1	40	40	1
Aspergillus/Penicillium - like	13	520	40	12	2	80	40	5	3	120	40	4
Basidiospores	73	2,920	40	67	35	1,400	40	80	75	3,000	40	91
Bipolaris/Dreschlera												
Chaetomium												
Cladosporium	9	360	40	8	7	280	40	16	2	80	40	2
Curvularia	1	40	40	1								
Epicoccum												
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces									1	40	40	1
Smuts/*Myxomycetes/Periconia*	2	80	40	2								
Stachybotrys												
Torula	2	80	40	2								
Trichoderma												
Ulocladium												
Other Spores												
Hyphal Fragments/ cm^2	3	120	40		2	80	40		3	120	40	
Total Fungal Spores/ m^3		4,360				1,760				3,280		

Annie Chang
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Katie Brown
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SELC Project #: **2025-3666**

Analysis Date: **12/29/25**
Report Date: **12/29/25**

Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**

PO Number: **None Given**

Lab ID # 100766

Sample Number	**5244**				**5197**				**5373**			
Sample Identification	Cell Block 1				Cell Block 2				Cell Block 3			
Sample Volume (liters)	75				75				75			
Debris Rating	2				3				2			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria					1	40	40	1				
Arthrinium												
Ascospores	7	280	40	11	5	200	40	4	4	160	40	5
Aspergillus/Penicillium - like	3	120	40	5	7	280	40	6				
Basidiospores	42	1,680	40	68	62	2,480	40	49	60	2,400	40	73
Bipolaris/Dreschlera												
Chaetomium												
Cladosporium	9	360	40	15	45	1,800	40	35	14	560	40	17
Curvularia					1	40	40	1	1	40	40	1
Epicoccum												
Fusarium												
Nigrospora					3	120	40	2				
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*					2	80	40	2	2	80	40	2
Stachybotrys	1	40	40	2								
Torula												
Trichoderma												
Ulocladium									1	40	40	1
Other Spores					1	40	40	1				
Hyphal Fragments/ cm^2	2	80	40		13	520	40		3	120	40	
Total Fungal Spores/ m^3		2,480				5,080				3,280		

Annie Chang

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421 Odell Ave.
Childersburg, Alabama
SELC Project #: **2025-3666**

Analysis Date: **12/29/25**
Report Date: **12/29/25**
Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**
PO Number: **None Given**

Lab ID # 100766

Sample Number	**5291**				**5305**				**5294**			
Sample Identification	Office Lobby				Unfinished Floor One				Unfinished Floor Two			
Sample Volume (liters)	75				75				75			
Debris Rating	1				1				3			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria												
Arthrinium												
Ascospores	2	80	40	5					16	640	40	10
Aspergillus/Penicillium - like									5	200	40	3
Basidiospores	37	1,480	40	95					102	4,080	40	66
Bipolaris/Dreschlera												
Chaetomium												
Cladosporium									10	400	40	6
Curvularia												
Epicoccum									2	80	40	1
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*									4	160	40	3
Stachybotrys									15	600	40	10
Torula												
Trichoderma												
Ulocladium												
Other Spores												
Hyphal Fragments/ cm^2	2	80	40						15	600	40	
Total Fungal Spores/ m^3		1,560			None Detected (<40)					6,160		

Annie Chang

Analyst
Annie Chang - Assistant Laboratory Director

Katie Brown

Technical Review
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Fungal Air Sample Analysis Report

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Lab ID # 100766

SELC

Customer: **A Plus Mold and Inspection Services**
421 Odell Ave.
Childersburg, Alabama

SELC Project #: **2025-3666**

Analysis Date: **12/29/25**
Report Date: **12/29/25**

Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**

PO Number: **None Given**

Sample Number	**5218**			
Sample Identification	HVAC Room			
Sample Volume (liters)	75			
Debris Rating	3			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria				
Arthrinium				
Ascospores	2	80	40	3
Aspergillus/Penicillium - like	3	120	40	4
Basidiospores	66	2,640	40	90
Bipolaris/Dreschlera				
Chaetomium				
Cladosporium	2	80	40	3
Curvularia				
Epicoccum				
Fusarium				
Nigrospora				
Oidium/Peronospora				
Pithomyces				
Smuts/*Myxomycetes/Periconia*				
Stachybotrys				
Torula				
Trichoderma				
Ulocladium				
Other Spores				
Hyphal Fragments/ cm^2	2	80	40	
Total Fungal Spores/ m^3		2,920		

Annie Chang

Analyst
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SELC

Fungal Swab Sample Analysis Report

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Phone: (205) 823-6200
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Lab ID # 100766

Customer: **A Plus Mold and Inspection Services**
421 Odell Ave.
Childersburg, Alabama

Analysis Date: **12/29/25**
Report Date: **12/29/25**

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**

SELC Project #: **2025-3666**

PO Number: **None Given**

Sample Number	**001**				**002**				**003**			
Sample Identification	Security Camera Room				Medical Room				Kitchen			
Sample Area (cm^2)	6.45				6.45				6.45			
Fungal Spore Identification	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**
Alternaria												
Arthrinium												
Ascospores									1	70	70	10
Aspergillus/Penicillium - like	280	26,040	93	99	96	24,960	260	100	2	140	70	20
Basidiospores	1	93	93	<1								
Bipolaris/Dreschlera												
Chaetomium												
Cladosporium									7	490	70	70
Curvularia	1	93	93	<1								
Epicoccum												
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*												
Stachybotrys												
Torula												
Trichoderma												
Ulocladium												
Other Spores												
Hyphal Fragments/ cm^2	8	744	93		4	1,040	260		2	140	70	
Total Fungal Spores/ cm^2		26,226				24,960				700		

Annie Chang

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Technical Review

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Safety Environmental Laboratories and Consulting, Inc.

SELC

Fungal Swab Sample Analysis Report

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Accredited Laboratory
Lab ID # 100766

Customer: **A Plus Mold and Inspection Services**
421 Odell Ave.
Childersburg, Alabama

Analysis Date: **12/29/25**
Report Date: **12/29/25**

Project Name: **Autauga County Jail**
Project Location: **Prattville, Alabama**

SELC Project #: **2025-3666**

PO Number: **None Given**

Sample Number	**004**				**005**			
Sample Identification	HVAC 1				Top Floor Mold On Box			
Sample Area (cm^2)	6.45				6.45			
Fungal Spore Identification	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**
Alternaria								
Arthrinium								
Ascospores	1	93	93	<1	1	93	93	<1
Aspergillus/Penicillium - like					39	3,627	93	10
Basidiospores								
Bipolaris/Dreschlera								
Chaetomium					2	186	93	<1
Cladosporium	287	26,691	93	100	357	33,201	93	87
Curvularia								
Epicoccum								
Fusarium								
Nigrospora								
Oidium/Peronospora								
Pithomyces								
Smuts/*Myxomycetes/Periconia*								
Stachybotrys					11	1,023	93	3
Torula								
Trichoderma								
Ulocladium								
Other Spores								
Hyphal Fragments/ cm^2	9	837	93		37	3,441	93	
Total Fungal Spores/ cm^2		26,784				38,130		

Annie Chang

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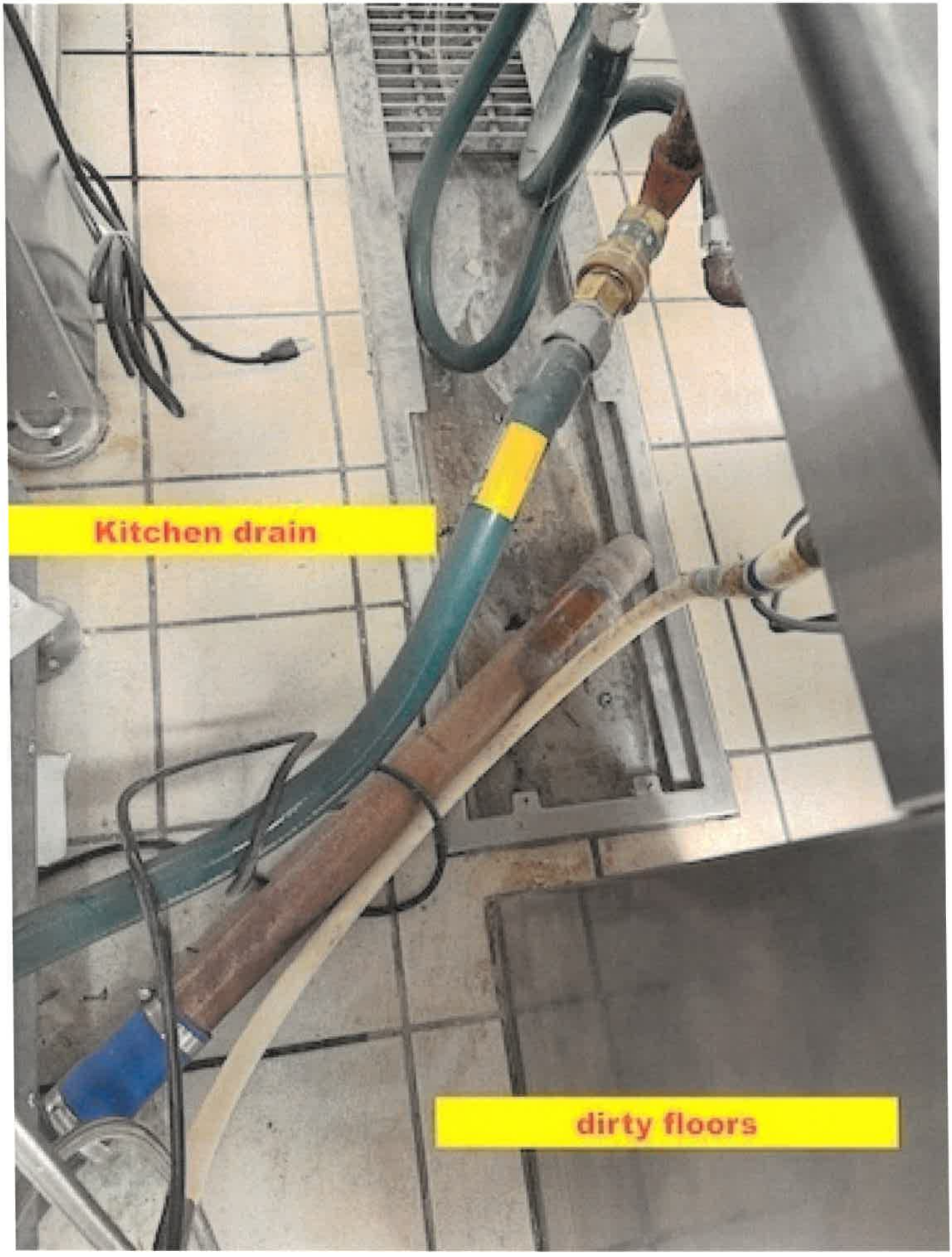
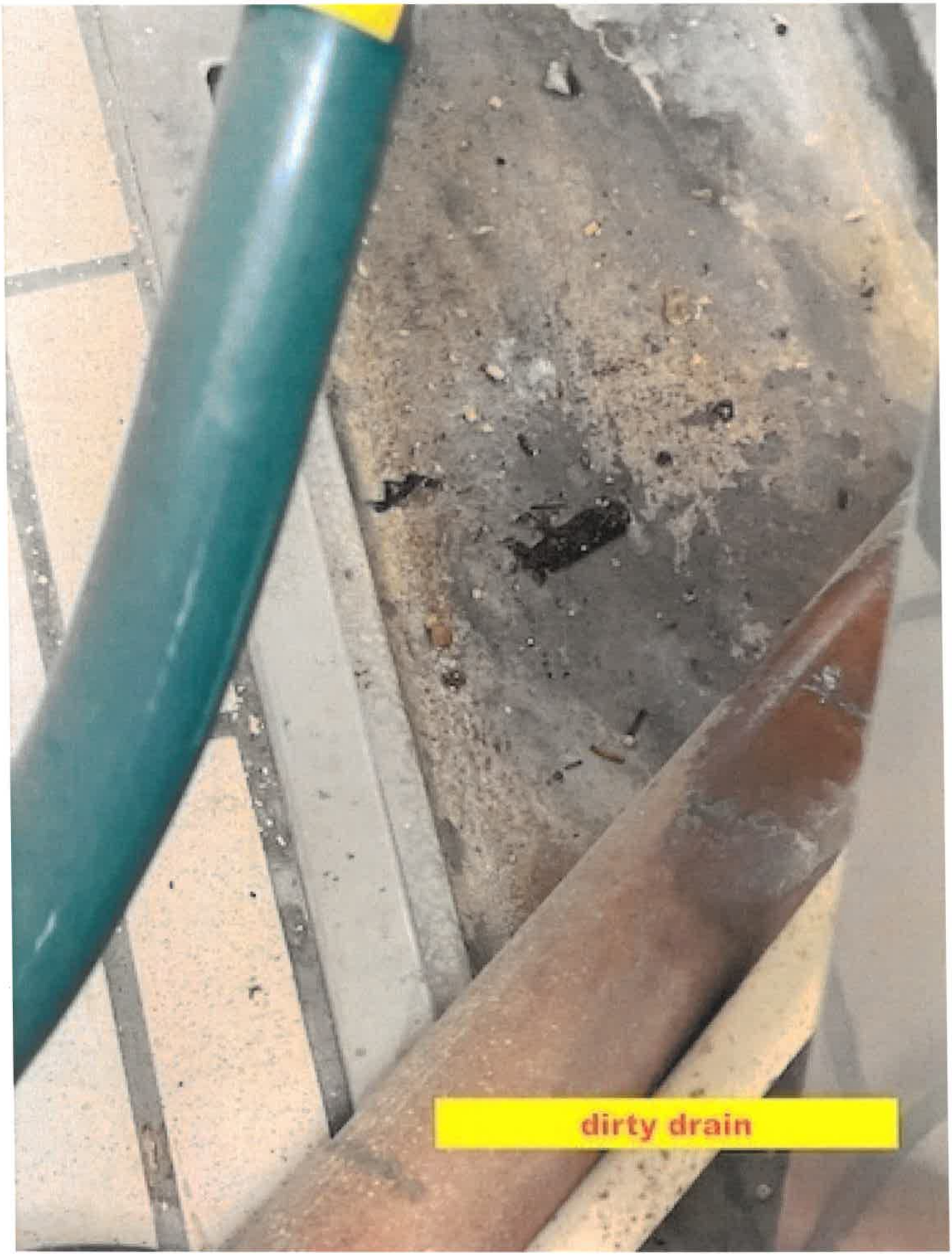
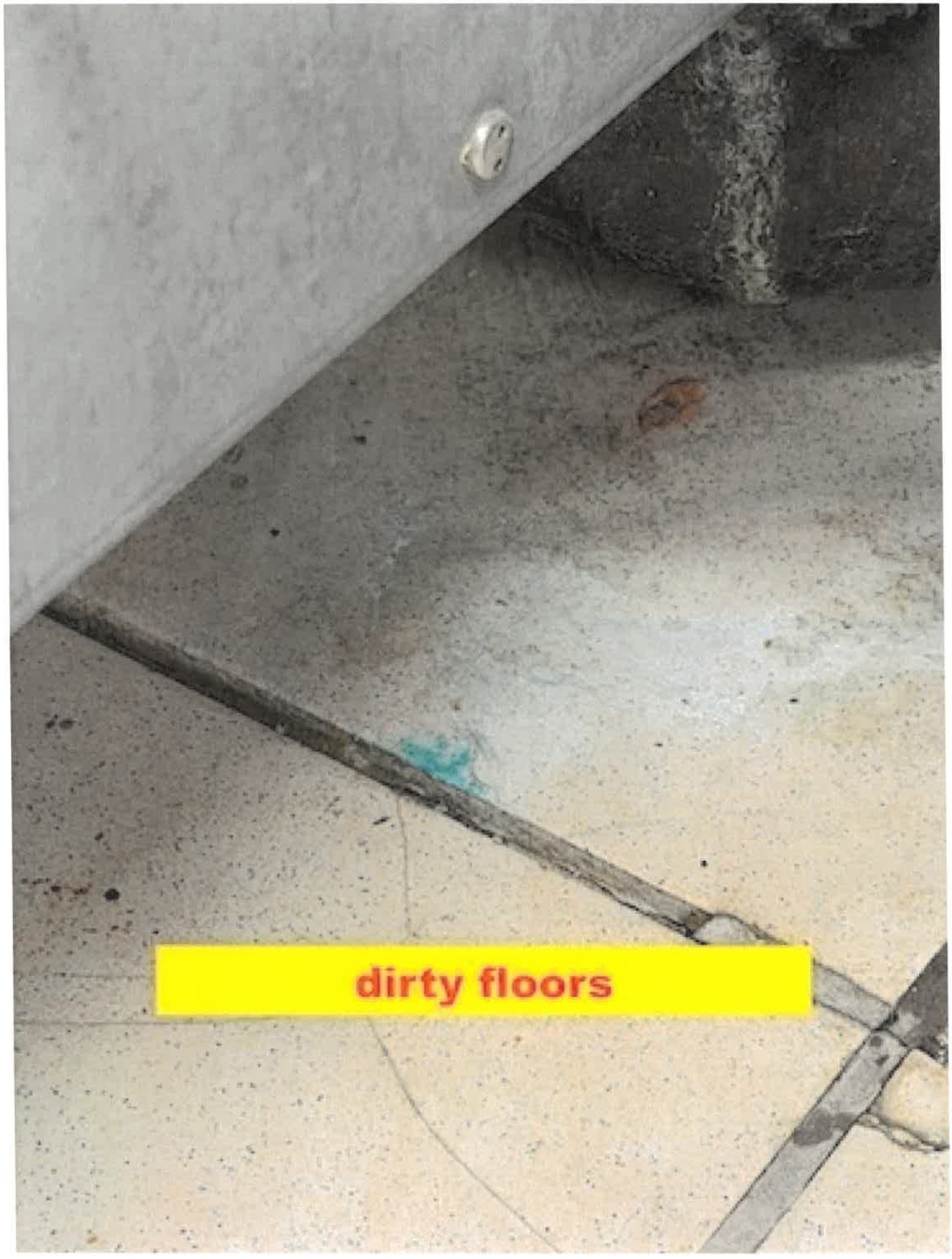

Kitchen drain
dirty floors


dirty drain

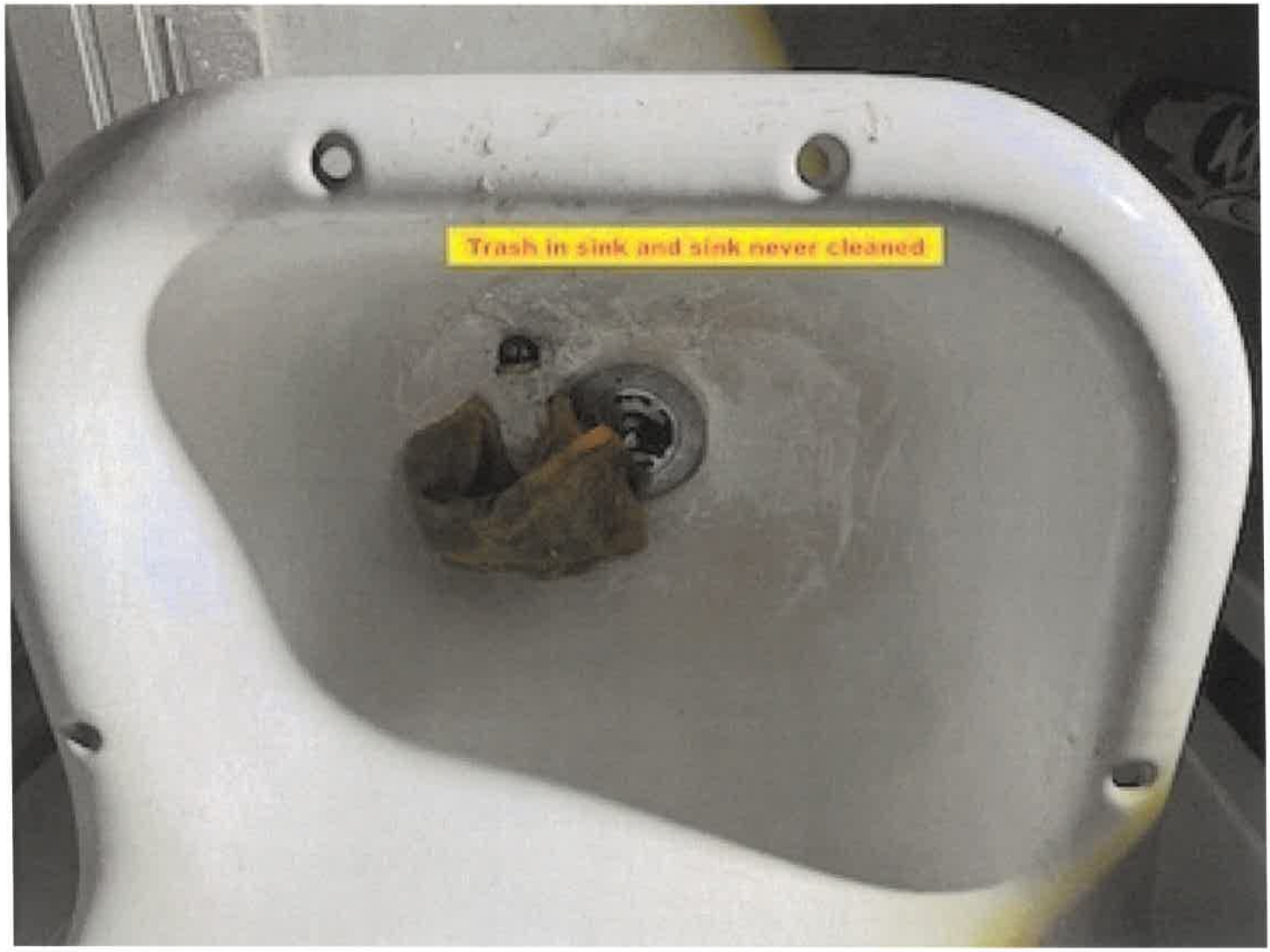

dirty floors


Dirty floors


Dirty floors

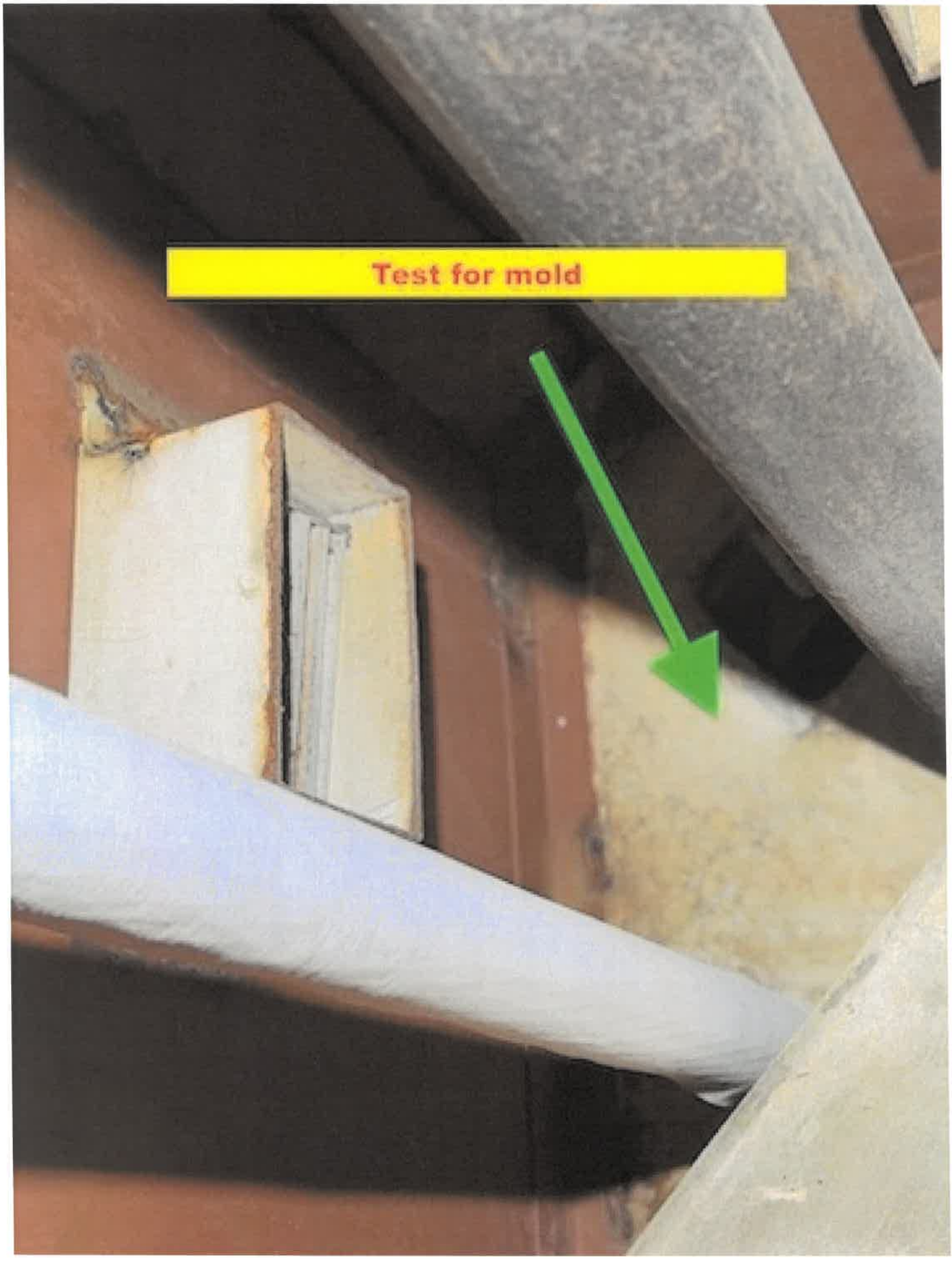

Dirty floors

Trash piled up

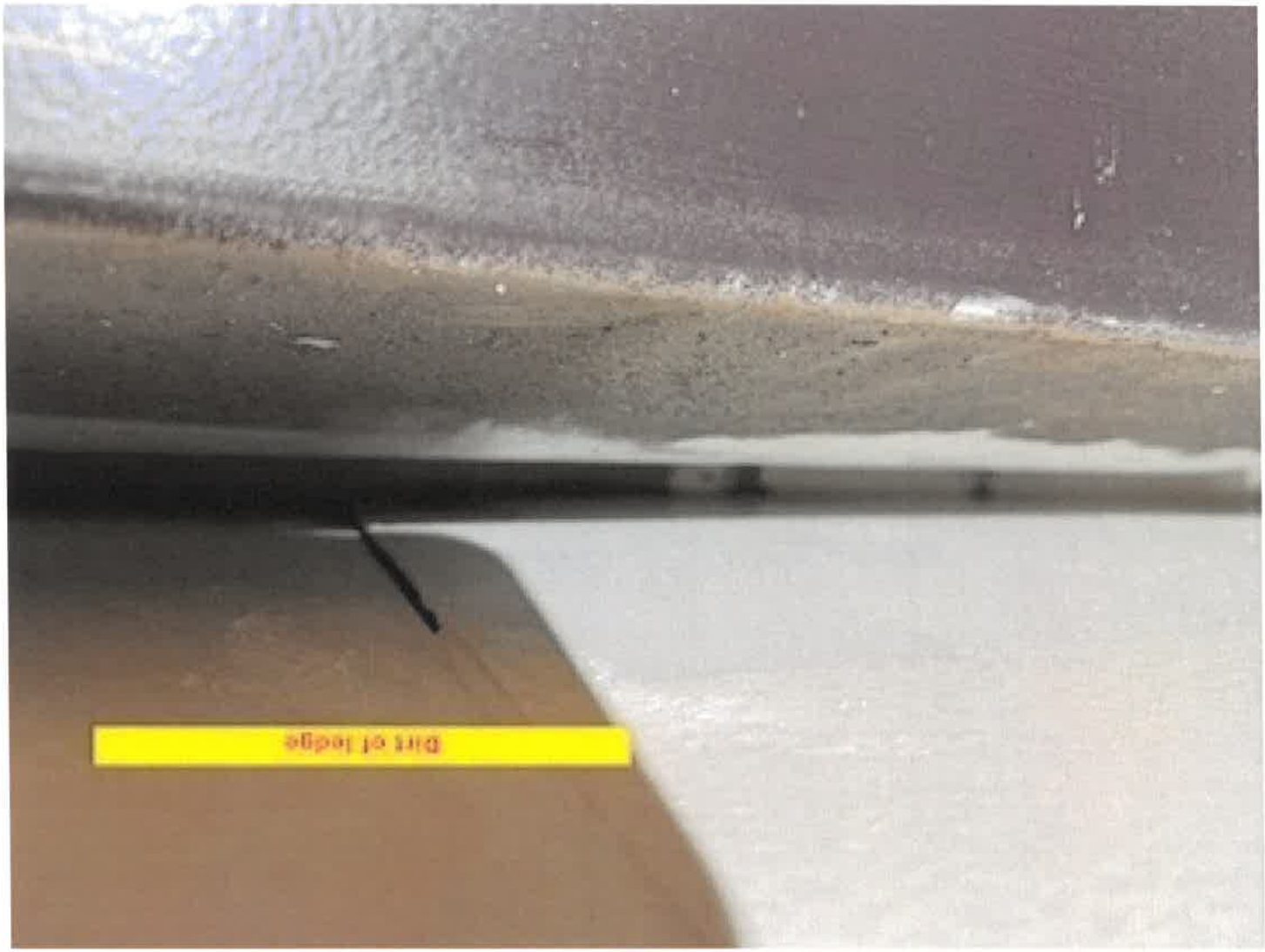

Trash in sink and sink never cleaned


Inside air return


Dirt inside HVAC system

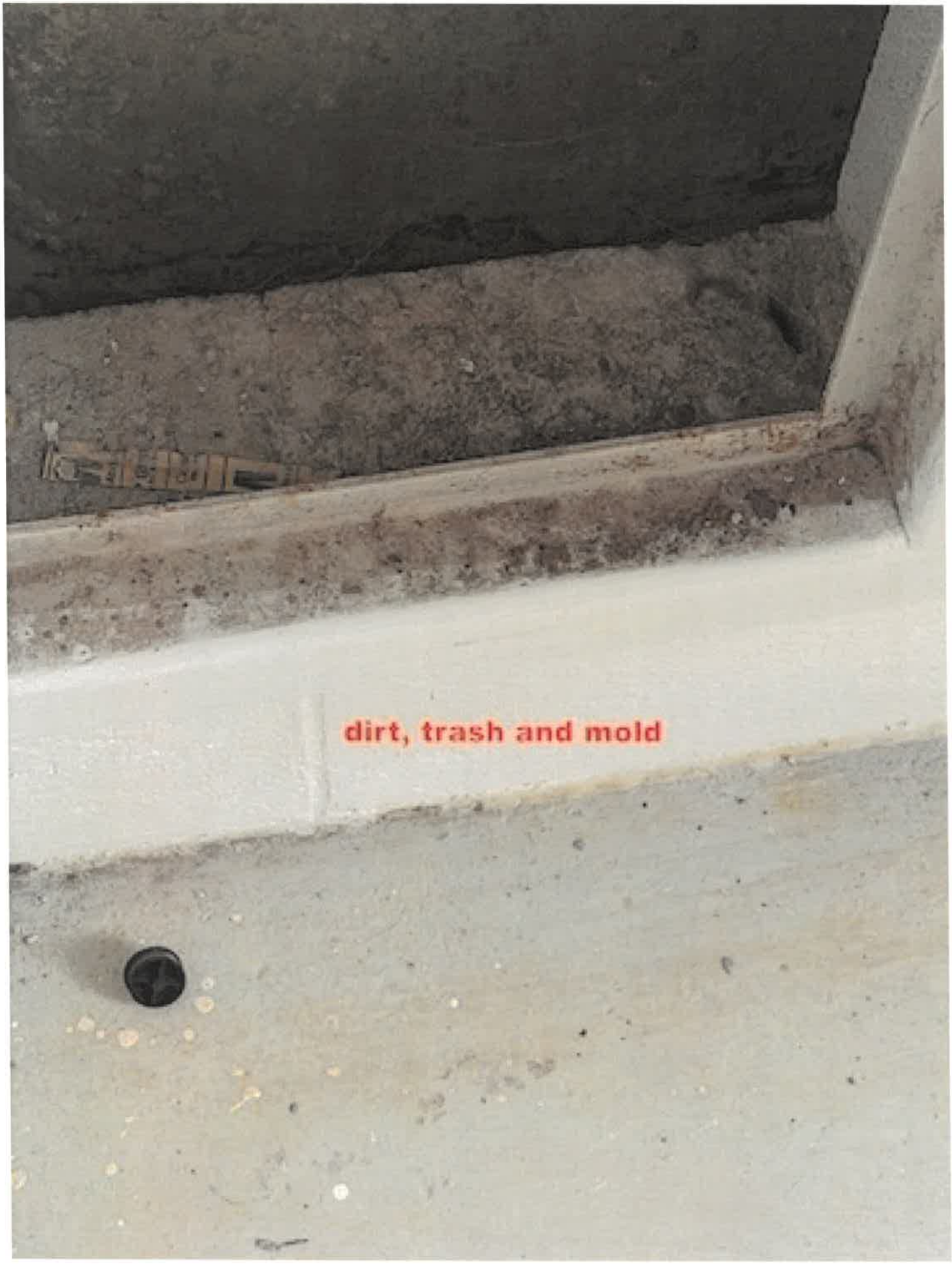

Test for mold


Dirty floors and side of metal

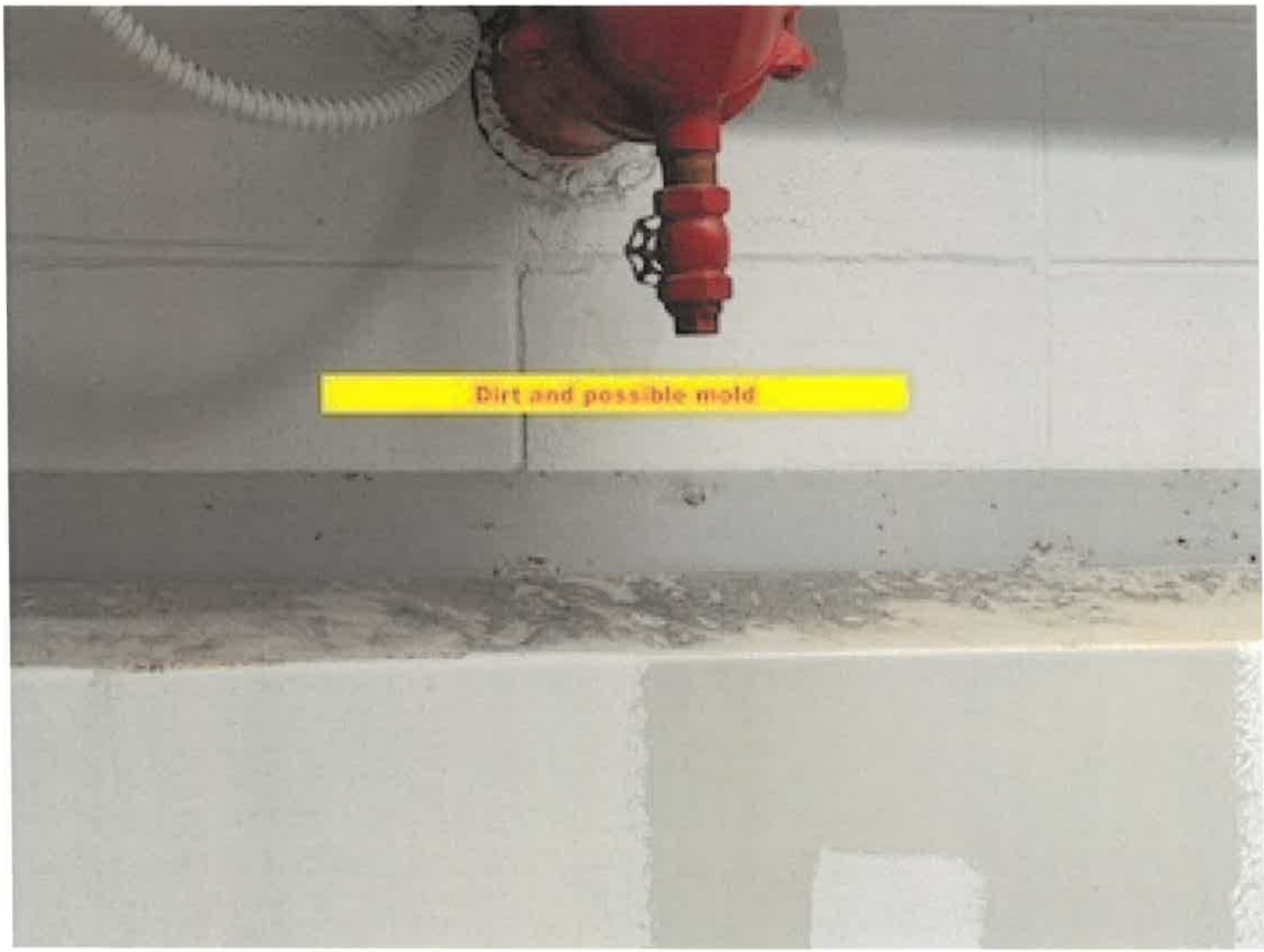

Dirt of lodge


Dirty on ledge


Dust and dirt on metal

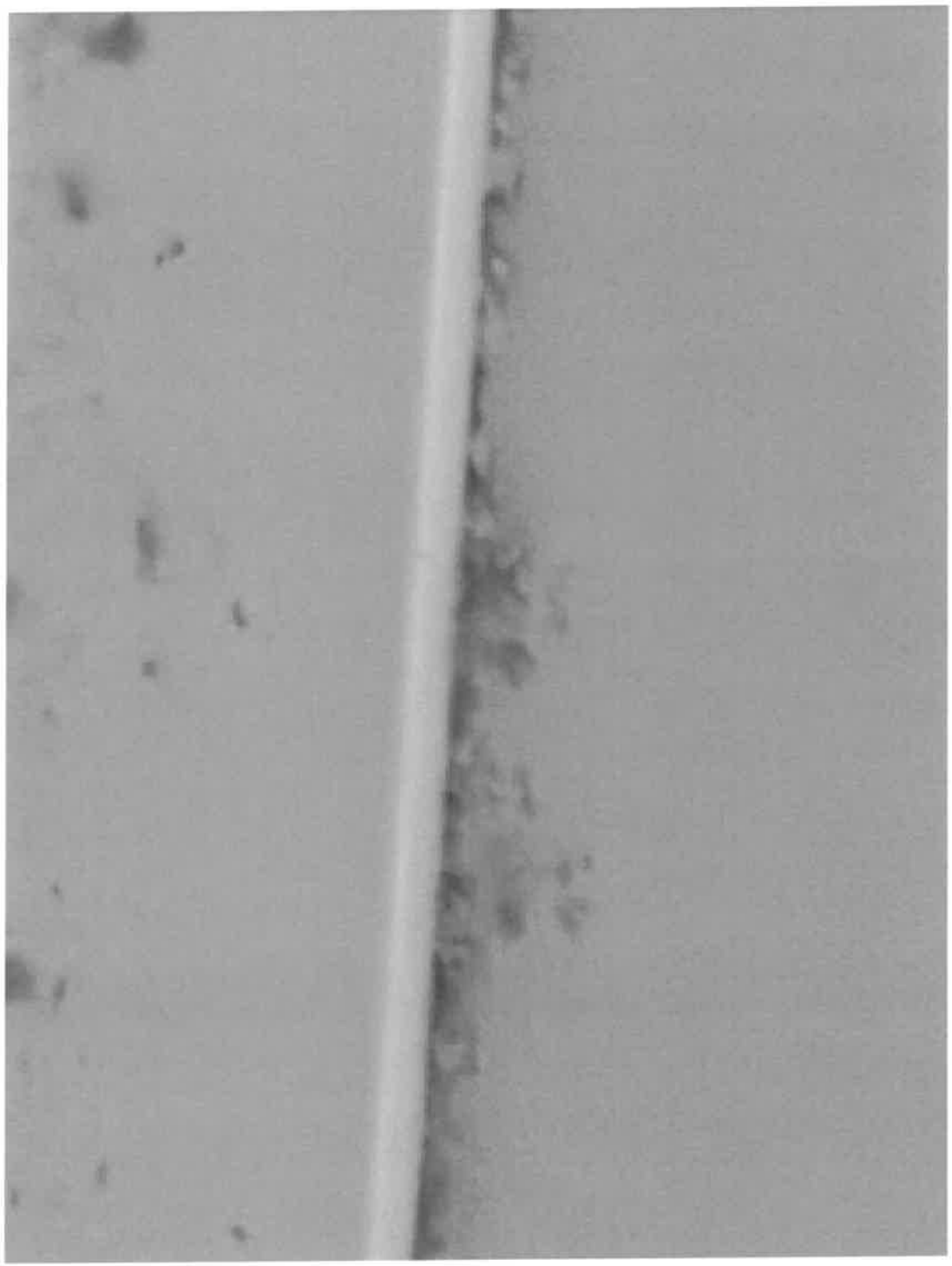

dirt, trash and mold


Rust and possible mold


Dirt and possible mold


Mold on stairs


Mold




Trash in electrical box and no work has been done in the this room.

New ductwork with heavy dust and contaminants
BELLY BAND HEIGHT COLOR
TRT – MOTOR
HARNESS – ECM
TRT – REVFLOW/SWITCH
TRT – REVFLOW/SWITCH
WHEEL

LIMITED INDOOR ENVIRONMENTAL (FUNGAL) ASSESSMENT AND REMEDIATION PROTOCOL

#2

OF THE

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA

SELC PROJECT # 2025-3659

SELC

Prepared for:

Autauga County Sherriff's Department
Attn: Sherriff Mark Harrell
162 West 4th Street
Prattville, AL 36067
mark.harrell@autauga.com

Prepared by:

Safety Environmental Laboratories and Consulting Inc.
P. O. Box 1848
Pelham, AL 35124
(205) 823-6200

December 2025

Safety Environmental Laboratories and Consulting, Inc.

P. O. Box 1848
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Phone: 205-823-6200
Fax: 205-823-9066

Environmental, Health, and Safety Solutions

January 5, 2026

Autauga County Sherriff's Department
Attn: Sherriff Mark Harrell
162 W 4th Street
Prattville, Alabama 36067

RE: Limited Indoor Environmental (Fungal/Mold) Assessment and Remediation Protocol of the Autauga County Jail Located at 136 N Court St., Prattville, Alabama 36067 (SELC Project # 2025-3659)

Sherriff Harrell:

On December 29, 2025, representatives of Safety Environmental Laboratories and Consulting, Inc (SELC) conducted a Limited Indoor Environmental (Fungal/Mold) Assessment of the Autauga County Jail located at 136 N Court St., Prattville, Alabama. The assessment was requested as a follow up to the initial inspection conducted in July 2024. The assessment was conducted in accordance with applicable and appropriate standard practices and guidelines put forth in *"Recognition, Evaluation, and Control of Indoor Mold"* by the American Industrial Hygiene Association and ASTM Standard D7338–14 (2023) *Standard Guide for the Assessment of Fungal Growth in Buildings* to evaluate the extent of fungal contamination within the structure and to provide remediation recommendations if necessary.

The scope of work included a thorough visual inspection and the collection of airborne and surface wipe fungal spore samples to characterize the potential fungal impact within the structure. Photographic documentation, building material moisture measurements, temperature, and humidity data were also collected.

It is important to note that the fungal remediation efforts conducted within the structure reportedly had been completed in October 2024. Construction efforts were ongoing at the time of the assessment. Workers were performing metal fabrication and modifications to the jail structure at the time of the assessment. In addition, some workers were painting and conducting general cleaning efforts at the time of the assessment.

The visual assessment of structure revealed moderate dust and debris accumulation on hard surfaces throughout the facility, specifically on horizontal surfaces such as exit signs, speaker boxes, the tops of the cell door support structure, tables, benches, shelving, cabinetry, and various other horizontal surfaces throughout the facility. A ceiling tile in the shift manager's office was visibly water stained. However, there was no evidence of significant water intrusion into the facility at the time of the assessment. An area of limited and localized fungal growth was observed on an exposed masonry block wall in the control room. There was no evidence of additional significant fungal contamination observed within the facility at the time of the assessment.

It is important to note that all heating, ventilation, and air conditioning (HVAC) systems and all associated supply and return ductwork serving the structure were replaced as part of the ongoing renovation efforts within the facility. Light to moderate particulate accumulation was observed in the newly installed HVAC system return and supply ductwork. In addition, the fresh air makeup ductwork was not replaced. Significant particulate accumulation was observed within the fresh air makeup ductwork serving the structure.

The visual inspection of the unfinished portion of the structure, specifically the third and fourth floor unfinished jail cell areas, revealed moderate dust accumulation on the majority of hard surfaces. It is important to note that there was no HVAC control or active dehumidification within the unfinished portion of the structure.

SECTION 1 BACKGROUND INFORMATION:

It should be noted that fungal spores are found within the air and on surfaces of all structures, including homes, schools, health care facilities, industrial and manufacturing facilities. Fungal organisms are part of our natural environment. However, elevated concentrations of water indicator and potentially toxigenic fungal forms are known to adversely impact the indoor environment.

While no known state or federal regulations have established permissible exposure limits for fungal contamination, in the event of a complaint of mold contamination in a place of employment, the Occupational Safety and Health Administration (OSHA) does frequently reference the General Duty Clause, Section 5(a)(1), which states that "each employer shall furnish to each of his employees employment and a place of employment which are free from recognized hazards that are causing or are likely to cause death or serious physical harm to his employees."

In addition, the U.S. Environmental Protection Agency (EPA) document titled *Moisture Control Guidance for Building Design, Construction and Maintenance*, December 2013, references a U.S. Centers for Disease Control and Prevention (CDC) and Institute of Medicine (IOM) committee finding for "damp or moldy indoor environments" that states, research shows an "association between exposure to damp indoor environments and adverse health effects, including: upper respiratory symptoms, cough, wheeze, asthma

symptoms in sensitized persons with asthma, dyspnea, lower respiratory illness in otherwise healthy children, and asthma development". Additionally, an increased risk for opportunistic infections in immune-compromised individuals also exists.

Typically, baseline / background airborne concentrations of fungal forms within residential and commercial facilities of the Southeastern United States will be similar to or below outdoor fungal spore concentrations and be composed of common environmental fungal forms. Furthermore, surfaces within these structures typically reveal a range from 10 to 500 fungal spores per square centimeter of surface area.

Surface samples collected from unconditioned spaces, areas of high air flow, and areas of significant dust accumulation such as attic spaces, crawlspaces, fresh air intakes, and HVAC components may reveal significantly higher spore surface densities depending on dust loading and age.

It should be noted that significantly contaminated or colonized surfaces, impacted by water intrusion events or long-term humidity issues may reveal mold spore surface densities ranging from tens of thousands (10,000's) to millions (1,000,000's) of spores per square centimeter of surface area. Likewise, mold spore concentrations in the air of significantly contaminated structures may reveal concentrations ranging from tens of thousands (10,000's) to millions (1,000,000's) of spores per cubic meter of air. Also, the predominant fungal (mold) spore compositions, found in such situations are not common environmental forms, but the water indicator and potentially toxigenic forms as designated by most laboratories. (EMLab RG)

SECTION 2 ANALYTICAL DATA:

The airborne fungal spore samples collected from pod 2, pod 3, pod 6, pod 7, the kitchen, the drug detection room, property room N113, and room 109A recovered concentrations ranging from 2,120 to 11,960 spores per cubic meter (spores/m^3). All eight indoor air samples recovered Basidiospores and/or *Cladosporium* as the predominant fungal forms identified.

The outdoor ambient air sample recovered 15,560 spores/m^3 with Basidiospores and *Cladosporium* as the predominant fungal forms identified. All eight indoor air samples recovered concentrations less the outdoors with similar predominant organisms. However, slightly elevated concentrations of *Aspergillus/Penicillium* group organisms were recovered from air samples collected from pod 2 and pod 3 when compared to the outdoors. In addition, lesser concentrations of *Chaetomium* were recovered from the pod 3 and drug detection room air samples, which was not recovered outdoors. Lesser concentrations of *Stachybotrys* were also recovered from the air sample collected from the drug detection room, which was not recovered outdoors.

The surface wipe sample collected from the control room exposed masonry block wall recovered 6,720 spores per square centimeter (spores/cm^2) with *Aspergillus/Penicillium*

group organisms and *Cladosporium* as the predominant fungal forms. The surface wipe sample collected from the HVAC fresh air intake ductwork recovered 5,040 spores/cm^2 with Basidiospores and *Cladosporium* as the predominant fungal forms identified. Lesser concentrations of *Aspergillus/Penicillium* group organisms and *Chaetomium* were also recovered from the fresh air intake ductwork surface sample. Both surface wipe samples are considered elevated when compared to typical background levels.

The composite surface wipe sample collected from room N113 records and property storage shelving recovered 27 spores/cm^2 with Basidiospores and *Chaetomium* as the only fungal forms identified. The composite surface wipe sample collected from the freezer located in the kitchen recovered 16 spores/cm^2 with Basidiospores as the only fungal form identified. The composite surface wipe sample collected from horizontal surfaces within the cell areas recovered 24 spores/cm^2 with Ascospores, *Aspergillus/Penicillium* group organisms, and *Ganoderma* as the only fungal forms identified. The composite surface wipe sample collected from the HVAC supply ductwork labeled C3 recovered 56 spores/cm^2 with Basidiospores, Ascospores, *Cladosporium*, and *Ganoderma* as the only fungal forms identified. All four aforementioned surface samples are considered to be at or below typical background concentrations.

Aspergillus/Penicillium group organisms, *Chaetomium*, and *Stachybotrys* are typically grouped as water indicator and potentially toxigenic fungal forms by most analytical laboratories. They are also classified as Type I and/or Type III allergens, causing hay fever, asthma and hypersensitivity pneumonitis in susceptible individuals (EMLAB RG).

Although *Cladosporium* is considered a common environmental fungal form, it is also largely recognized as a very important allergen by most analytical laboratories. Elevated concentrations of *Cladosporium* are often indicative of excessive particulate / dust accumulation, but may also indicate mold growth.

Moisture measurements collected from interior finishing materials ranged from 5% to 15%. Wood science studies have shown that moisture concentrations at or above 20% to 28% are adequate to sustain microbial growth.

Temperature and relative humidity measurements within the structure ranged from 67.1° to 73.7° Fahrenheit and 19.8% to 23.3% respectively. Guidance provided by the American Society of Refrigeration, Heating and Air Conditioning Engineers (ASHRAE) and the U.S. EPA, indicate that extended periods of relative humidity greater than 60% can result in microbial proliferation. These organizations also recommend a comfort level of 30% to 50% for most indoor environments. The relative humidity outdoors was 29.3% at the time of the assessment.

SECTION 3 CONCLUSIONS:

Based upon the analytical data and the visual inspection, limited and localized fungal growth is impacting a portion of the exposed masonry block wall in the control room. Heavy particulate accumulation containing slightly elevated concentrations of potentially toxigenic and water indicator fungal forms are also impacting the fresh air intake ductwork serving the facility. However, it is important to note that the fungal contamination on surfaces within the control room and the fresh air intake do not appear to be significantly impacting the air quality within the facility at this time. The slightly elevated potentially toxigenic and water indicator fungal spore concentrations recovered from pod 2, pod 3, and the drug detection room are likely resulting from the significant construction dust and debris observed on surfaces throughout the facility.

The limited fungal contamination on portions of the control room masonry block wall appears to be a result previous condensation issues prior to HVAC replacement efforts. It is important to note that the visible fungal growth on the control room masonry block wall appears to be less than ten square feet (<10 ft^2). Therefore, the remediation efforts in the control room are determined to be small by EPA guidelines and may be conducted by the property owner or a general contractor. It is recommended to ensure the HVAC systems are utilized to maintain indoor relative humidity within the structure to less than 60% at all times.

Following this report, you will find a recommended remediation protocol. Attached you will find Appendix A which presents the analytical data in table form, Appendix B which contains the photographic documentation, Appendix C which contains the analytical laboratory reports, and Appendix D which contains supplemental fungal information.

Please be advised that these recommendations do not imply coverage of these issues by the insurance carrier. They are presented as guidance to support the remediation and restoration effort as it relates to indoor environmental quality.

Safety Environmental Laboratories and Consulting, Inc. is pleased to have provided you with these services. If you have any questions concerning this information, please feel free to contact me at (205) 823-6200.

Sincerely,

Brad Stiles, CIEC
Director of Operations / Senior Project Manager
Certified Indoor Environmental Consultant #1203015

Reviewed By

Neil Matson, CIH, CIEC, CHMM, REM, REPA
Vice President of Operations
Certified Industrial Hygienist #11266
Certified Indoor Environmental Consultant #1202015
Certified Hazardous Materials Manager #16192
Registered Environmental Manager #12554
Registered Environmental Property Assessor #512867636

Safety Environmental Laboratories and Consulting, Inc.

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Pelham, AL 35124

Phone: 205-823-6200
Fax: 205-823-9066

Environmental, Health, and Safety Solutions

December 29, 2025

Autauga County Jail
136 N Court St.
Prattville, Alabama 36067
(SELC Project # 2025-3659)

RECOMMENDED REMEDIATION AND RESTORATION PROTOCOL

1. It is recommended to ensure the HVAC systems serving the structure are utilized to maintain indoor relative humidity within the structure to less than 60% at all times.
2. Additional HVAC usage or supplemental dehumidification may be necessary if humidity levels exceed 60% within the unfinished portion of the structure, specifically on floors 3 and 4.
3. It is recommended that personnel conducting all removal and/or cleaning efforts of visibly contaminated building materials wear appropriate personal protective equipment. PPE should include, at a minimum, a N-95 respirator, disposable clothing, eye protection, and disposable gloves.

Control Room North Block Wall

4. The areas exhibiting visible fungal growth in the control room appear to be less than 10 square feet of continuous fungal growth in size. Therefore, the use of negative pressure containments does not appear to be necessary. However, it is recommended that HEPA filtration units set to recycle mode be utilized throughout the control room during remediation activities to reduce airborne fungal spore concentrations.
5. The computer systems and monitors should either be removed from the control room or covered and sealed in polyethylene sheeting prior to conducting block wall cleaning efforts.
6. The visibly impacted portions of the masonry block wall along the north wall of the control room should be cleaned utilizing hand cleaning techniques with a suitable anti-microbial solution or a mild detergent and water solution. Cleaning efforts should extend two feet beyond the point of visible fungal growth.
7. Hand cleaning techniques should focus on removal of fungal spores from surfaces rather than killing the spores. A clean cloth should be used, wiping in a continuous motion until the cloth becomes soiled; at which time, the cloth should be discarded.

Fresh Air Intake Ductwork

8. It is recommended that all fresh air intake ductwork serving the structure be cleaned by a professional contractor experienced in cleaning of HVAC systems for mold contamination. Cleaning of the HVAC system fresh air intake ductwork should be conducted in accordance with the National Air Duct Cleaners Association (NADCA) guidelines.

General Guidance

9. Following completion of construction/renovation efforts within the facility, it is recommended to conduct routine housekeeping / post construction cleaning efforts throughout the facility. Cleaning efforts should focus on removing dust and debris from horizontal surfaces throughout the facility. This should include exit signs, speaker boxes, the tops of cell door support structure, tables, benches, shelving, cabinetry, hard surface flooring, window and door framing, stairs, handrailing, fixtures, and any other horizontal surfaces exhibiting evidence of construction dust and debris accumulation throughout the facility.
10. Following completion of control room cleaning and fresh air intake cleaning efforts, it is recommended to filter the air for a period of 48 to 72 hours utilizing high efficiency particulate air (HEPA) filtration units. This will allow any airborne fungal spores generated during the work activities to be filtered through the HEPA units. This should include, at a minimum, utilizing HEPA filtration systems in the control room, pod 2, pod 3, and the drug detection room.
11. Upon completion of the limited remediation and cleaning efforts, no further testing for fungal contamination is determined to be warranted at this time.

APPENDIX A

TABLES

TABLE A – AIRBORNE FUNGAL SPORE ANALYSIS

TABLE B – SURFACE WIPE FUNGAL ANALYSIS

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA
DECEMBER 29, 2025

TABLE A: TOTAL AIRBORNE FUNGAL SPORE ANALYSES

SAMPLE LOCATION	CONCENTRATION[1], spores/m^3	IDENTIFICATION[2]
#1 - 6 Pod	10,440	Basidiospores (56%) Cladosporium (31%) Ascospores (9%) Aspergillus/Penicillium (2%) Smuts/Myxomycetes/Periconia (1%) Alternaria (<1%) Nigrospora (<1%) Other Spores (<1%)
#2 - 2 Pod	8,600	Basidiospores (47%) Cladosporium (32%) Aspergillus/Penicillium (17%) Ascospores (4%) Smuts/Myxomycetes/Periconia (1%) Alternaria (<1%)

1-Concentration is measured in spores per cubic meter of air, as determined from impaction onto a spore trap, collected with a high volume pump, set to 15 liters per minute for ten minutes, totaling 150 liters per sample.

2-Species identified are listed according to predominance.

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA
DECEMBER 29, 2025

TABLE A: TOTAL AIRBORNE FUNGAL SPORE ANALYSES

SAMPLE LOCATION	CONCENTRATION[1], spores/m^3	IDENTIFICATION[2]
#3 - 3 Pod	9,600	Basidiospores (45%) Aspergillus/Penicillium (23%) Cladosporium (23%) Ascospores (7%) Chaetomium (<1%) Epicoccum (<1%) Smuts/Myxomycetes/Periconia (<1%)
#4 - 7 Pod	11,960	Basidiospores (65%) Cladosporium (29%) Ascospores (4%) Aspergillus/Penicillium (2%) Alternaria (1%)
#5 - Kitchen	4,280	Basidiospores (53%) Cladosporium (36%) Aspergillus/Penicillium (6%) Ascospores (5%) Smuts/Myxomycetes/Periconia (1%)

1-Concentration is measured in spores per cubic meter of air, as determined from impaction onto a spore trap, collected with a high volume pump, set to 15 liters per minute for ten minutes, totaling 150 liters per sample.

2-Species identified are listed according to predominance.

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA
DECEMBER 29, 2025

TABLE A: TOTAL AIRBORNE FUNGAL SPORE ANALYSES

SAMPLE LOCATION	CONCENTRATION[1], spores/m^3	IDENTIFICATION[2]
#6 - Drug Detection Room	10,720	Cladosporium (50%) Basidiospores (40%) Aspergillus/Penicillium (5%) Ascospores (3%) Stachybotrys (2%) Chaetomium (1%)
#7 - Room N113	3,720	Basidiospores (74%) Aspergillus/Penicillium (11%) Cladosporium (11%) Ascospores (3%) Smuts/Myxomycetes/Periconia (1%)
#8 - Room 109A	2,120	Basidiospores (92%) Cladosporium (4%) Ascospores (2%) Aspergillus/Penicillium (2%)

1-Concentration is measured in spores per cubic meter of air, as determined from impaction onto a spore trap, collected with a high volume pump, set to 15 liters per minute for ten minutes, totaling 150 liters per sample.

2-Species identified are listed according to predominance.

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA
DECEMBER 29, 2025

TABLE A: TOTAL AIRBORNE FUNGAL SPORE ANALYSES

SAMPLE LOCATION	CONCENTRATION[1], spores/m^3	IDENTIFICATION[2]	
#9 - Outside Ambient Air	15,560	Basidiospores	(66%)
		Cladosporium	(19%)
		Ascospores	(7%)
		Aspergillus/Penicillium	(6%)
		Alternaria	(1%)
		Smuts/Myxomycetes/Periconia	(1%)
		Epicoccum	(<1%)
		Ulocladium	(<1%)

1-Concentration is measured in spores per cubic meter of air, as determined from impaction onto a spore trap, collected with a high volume pump, set to 15 liters per minute for ten minutes, totaling 150 liters per sample.

2-Species identified are listed according to predominance.

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA
DECEMBER 29, 2025

TABLE B: SURFACE WIPE FUNGAL ANALYSES

SAMPLE LOCATION	CONCENTRATION[1], spores/cm^2	IDENTIFICATION[2]
#S1 - Control Room Wall	6,720	Aspergillus/Penicillium (68%) Cladosporium (21%) Ascospores (7%) Curvularia (4%)
#S2 - Records Shelving	27	Basidiospores (67%) Chaetomium (33%)
#S3 - Freezer Wall	16	Basidiospores (100%)
#S4 - Cell Horizontal Surfaces	24	Ascospores (33%) Aspergillus/Penicillium (33%) Ganoderma (33%)

1-Collected with a moistened cotton swab over a 10 to 100 square centimeter area. Concentration is measured in spores per square centimeter (spores/cm^2)

2-Species identified are listed according to predominance.

AUTAUGA COUNTY JAIL
136 NORTH COURT STREET
PRATTVILLE, ALABAMA
DECEMBER 29, 2025

TABLE B: SURFACE WIPE FUNGAL ANALYSES

SAMPLE LOCATION	CONCENTRATION[1], spores/cm^2	IDENTIFICATION[2]	
#S5 - C3 Duct Supply	56	Basidiospores Ascospores Cladosporium Ganoderma	(43%) (29%) (14%) (14%)
#S6 - Fresh Air Intake for Cells	5,040	Basidiospores Cladosporium Aspergillus/Penicillium Ascospores Chaetomium	(40%) (33%) (19%) (5%) (2%)

1-Collected with a moistened cotton swab over a 10 to 100 square centimeter area. Concentration is measured in spores per square centimeter (spores/cm^2)

2-Species identified are listed according to predominance.

APPENDIX B

PHOTOGRAPHIC DOCUMENTATION

Photographic Documentation

6 Pod, General View

6 Pod, Location of Indoor Airborne Fungal Spore Sample

2 Pod, General View

2 Pod, Location of Indoor Airborne Fungal Spore Sample

SELC Project # 2025-3659

Autauga County Jail
136 N Court St.
Prattville, AL 36067

December 29, 2025

Photographic Documentation

3 Pod, General View

3 Pod, Location of Indoor Airborne Fungal Spore Sample

3 Pod, Particulate Accumulation on Metal Door

3 Pod, Particulate Accumulation on Metal Door

Photographic Documentation

7 Pod, General View

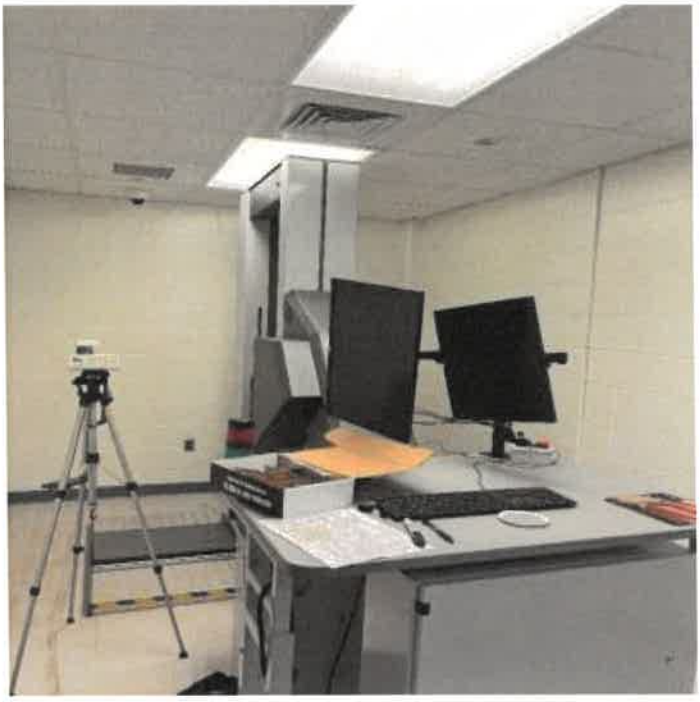
7 Pod, Location of Indoor Airborne Fungal Spore Sample

7 Pod, Particulate Accumulation on Metal Stool

Kitchen, General View

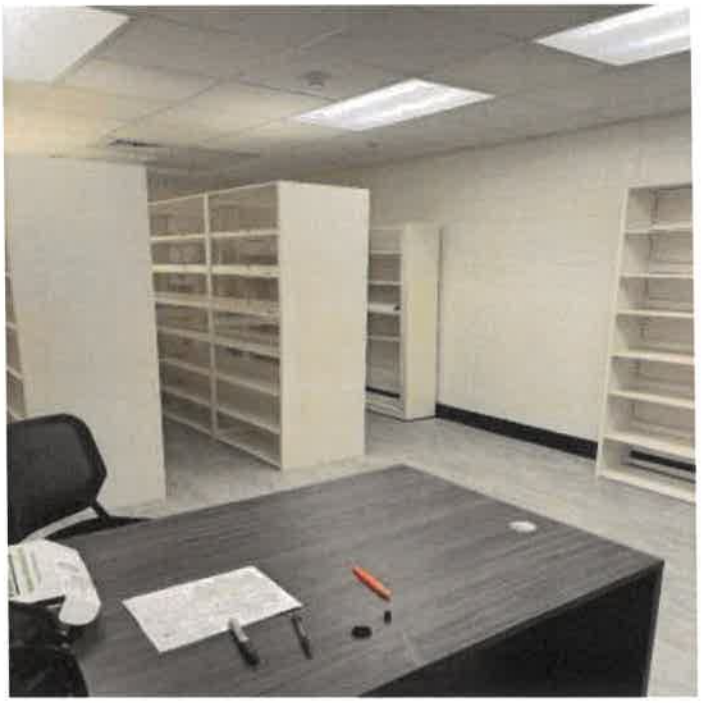
Photographic Documentation

Kitchen, Location of Indoor Airborne Fungal Spore Sample



Drug Detection Room, General View

Drug Detection Room, Location of Indoor Airborne Fungal Spore Sample



Room N113, General View

Photographic Documentation

Room N113, Location of Indoor Airborne Fungal Spore Sample

Room 109A, General View

Room 109A, Location of Indoor Airborne Fungal Spore Sample

Room 109A, Light Particulate Accumulation on Shelving

Photographic Documentation

Location of Outdoor Ambient Air Sample

APPENDIX C

ANALYTICAL RESULTS

AND

CHAIN OF CUSTODY

SELC

MICROBIAL LABORATORY FUNGAL ANALYSIS REPORT

Indoor Environmental (Mold/Fungal) Report

Prepared For:

SELC
989 Yeager Parkway
Pelham, AL 35124

Project Name: Autauga County Jail
Project Location: 136 N. Court St.
Prattville, AL 36067
PO Number: None Given

SELC Project #: **2025-3659**
Report Date: **12/30/2025**

Lab ID # 100766

MICROBIAL LABORATORY FUNGAL ANALYSIS REPORT:

REPORT USE AND DISCLAIMER:

Please be advised that all fungal (mold) assessments and inspections are required to have a thorough visual inspection of the structure with fungal (mold) sampling conducted only to support the Indoor Environmental Quality Professionals (IEQP) observations. This laboratory report alone doesn't constitute a fungal (mold) assessment or inspection as put forth in "*Recognition Evaluation, and Control of Indoor Mold*" by the American Industrial Hygiene Association (AIHA), or the ASTM Standard D7338-14 "*Standard Guide for the Assessment of Fungal Growth in Buildings*".

Furthermore, the AIHA states "*Prior to collecting any air samples for mold spores, the indoor environmental quality (IEQ) professional must determine the purpose and relevance of the sampling as well as ascertain the questions the sampling will answer. Air sampling should be considered as a screening tool or as ancillary to an informed inspection. Testing results should confirm observations or otherwise support conclusions made based on the informed inspection. In the absence of an informed inspection, air sampling alone cannot support any definitive conclusions. Air sampling for mold spores does not and cannot evaluate potential health risks.*" (AIHA, 2019)

SECTION 1 BACKGROUND INFORMATION:

1.1 General Information:

The following background information is provided to assist the client in evaluating the analytical results revealed by the mold / fungal analyses. This information is based on years of sampling and field experience, published references, governmental guidelines and association standards.

It should be noted that mold / fungal spores are found within the air and on surfaces of all structures, including homes, schools, health care facilities, industrial and manufacturing facilities. Fungal organisms are a part of our natural environment. However, elevated concentrations of water indicator and potentially toxigenic fungal forms are known to adversely impact the indoor environment.

At this time, there are no federal regulations governing the types or concentrations of molds / fungi for the indoor environment. A listing of pertinent references is provided later in this report.

1.2 Common Groupings of Mold / Fungal Forms:

References to "common environmental mold / fungal forms" has come to mean those fungal organisms routinely found in the outside / ambient air environment and therefore

the fungal forms we are exposed to on a daily basis. Examples of such genera or groupings are *Alternaria*, Ascospores, Basidiospores, *Cladosporium*, *Curvularia*, *Pithomyces*, Rusts, Smuts, etc.

References to "water indicator and potentially toxigenic fungal forms" have come to mean those fungal organisms, which have a high-water requirement and are known to produce various mycotoxins when subjected to certain environmental stimuli. Examples of such genera include *Aspergillus*, *Chaetomium*, *Fusarium*, *Penicillium* and *Stachybotrys*. This grouping contains the commonly referred to "toxic black molds".

1.3 Comparison of Background / Baseline Fungal Analyses:

The following comparisons may be helpful to the Client for understanding the analytical method of reporting spore concentrations and surface density units. Air concentrations are reported in total fungal spores per cubic meter of air (spores/m^3). A cubic meter is roughly equivalent to 35 cubic feet or 264 gallons. Tape lifts and surface wipes are reported in units of total spores per square centimeter (spores/cm^2). A square centimeter is roughly equivalent to the area of one face of a sugar cube. Bulk dust and building material samples are reported in units of total spores per gram of material (spores/g). There are 454 grams per pound or 28 grams per ounce.

Typically, baseline / background total airborne concentrations of fungal forms within residential and commercial facilities of the southeastern United States range between 100 to 1,000 spores per cubic meter of air and are composed of common environmental fungal forms found in the outdoor / ambient air.

Furthermore, hard surfaces, such as furniture and walls, within these structures typically reveal a baseline / background range of 1 to 100 fungal spores per square centimeter of surface area and are composed of common environmental forms. These values will vary depending on the thoroughness of housekeeping, humidity and moisture loadings within the structure.

The American Conference of Governmental Industrial Hygienist (ACGIH) provides the following guidance concerning data interpretation; "Data from individual sampling episodes is often interpreted with respect to baseline data from other environments or the same environment under anticipated low exposure conditions". In common terms, and with the absence of established acceptable exposure limits, it is often necessary to use a comparison value as a benchmark or "standard" when interpreting fungal (mold) data.

Also, ACGIH states that "differences that can be detected with manageable sample sizes are likely to be in 10-fold multiplicative steps (e.g., 100 versus 1,000)". This statement infers that if the total fungal spore count is ten (10) times greater in the sample from the suspect area than in the negative control sample collected from a non-suspect area (background / baseline), then that sample area may be a fungal

amplification site. ACGIH further states that "active fungal growth in indoor environments is inappropriate and may lead to exposure and adverse health effects."

SECTION 2 METHODOLOGY:

Safety Environmental Laboratories and Consulting, Inc. (SELC) provides direct microscopic analyses for fungal organisms on spore traps (air samples), surface wipes (sterile swabs), tape lifts, bulk dusts and bulk building materials.

Microscopic quantitative analyses are conducted at 400X to 600X magnification. Identification of fungal organisms may be further enhanced by the use of stains and oil immersion techniques at 1,000X magnification.

The results are reported as total fungal spores, meaning they include both viable and non-viable fungal spores. Direct microscopic techniques do not allow for the differentiation of *Aspergillus* and *Penicillium* spores. Therefore, such spores are reported as *Aspergillus / Penicillium* –like.

Also, depending on the spores morphology (shape, size, structure), other non-distinctive spores will be reported in groups or categories such as Ascospores, Basidiospores, or colorless / brown spore groups.

2.1 Air Samples – Spore Trap Analyses:

All analysis of spore trap media is conducted in accordance to ASTM published method D7391-20.

Typically, during the analysis of the sample thirty-three percent (33%) of the trace (collection area) of the spore trap (air sample) is analyzed. If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the trace and extrapolate the result for the entire sample. Fungal forms with greater than 500 spores per sample (an elevated fungal count) are difficult to analyze. Therefore, a sample with a significantly elevated fungal concentration may be an approximation.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3. The limit of detection is the analytical sensitivity (in spores/m^3) multiplied by the sample volume (in liters) divided by 1000 liters.

Excessive non-fungal particulate matter can obscure the presence of fungal spores. Such sample conditions can reduce the analytical accuracy. All samples are evaluated by the following debris scale for data qualification.

Air Sample Debris Rating Scale

Debris Rating	Description	Interpretation
0	No particulate detected	The absence of particulate matter on the slide could indicate improper sampling or a blank sample. Analyst should note accordingly.
1	Minimal particulate matter present. (0% to 5%)	Reported values are minimally affected by debris (typically 0-5% coverage of sample surface area)

2	Approx. 5% to 25% of trace occluded with particulate matter.	Negative bias is expected. The degree of bias increases with the percent of the trace that is occluded.
3	Approx. 25% to 75% of the trace occluded with particulate matter.	Negative bias is expected. The degree of bias increases with the percent of the trace that is occluded.
4	Approx. 75% to 90% of trace occluded with particulate matter.	Non-fungal particulate can mask fungal spores. Actual values may be moderately to significantly greater than the values reported. Negative bias is expected and increases with percentage of trace analyzed.
5	>90% of slide occluded with non-fungal debris.	Quantitative analysis is not possible. Sample trace is overloaded and cannot be reliably quantitatively analyzed due to excessive particulate matter. A new sample should be collected at shorter time interval, or other measures taken to reduce the particle load. Identification and an estimation of concentration may be provided at the Laboratory's discretion. The laboratory shall report presence only.

2.2 Surface Wipe Samples – Sterile Swabs:

Fungal spores and hyphae are extracted from the swab by mechanical action. The fungal components are placed on a slide, stained, identified and counted. The results are reported as spores per sample or spores per square centimeter, based upon the client supplied sample area.

If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the sample preparation and extrapolate the result for the entire sample. Therefore, a sample with a significantly elevated fungal count may be an approximation.

The limit of detection is 1 spore per area analyzed.

The analytical sensitivity is (1 spore/Total number of fields observed)* (Total Sample Area (cm^2)/Field Area of the microscope objective (cm^2))* 1/unit volume)*Dilution Factor.

2.3 Surface Samples – Tape Lifts:

All analysis of tape lift media is conducted in accordance to ASTM published method D 7658-17R21.

Fungal spores and hyphae are collected on the adhesive side of clear tape. The sample is placed on a slide, identified and counted. The results are reported as spores per square centimeter. If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the sample preparation and extrapolate the result for the

entire sample. Therefore, a sample with a significantly elevated fungal count may be an approximation.

2.4 Bulk Samples – Dusts and Building Materials:

The bulk sample is weighed and fungal components are extracted or lifted from the material. The fungal components are placed on a slide, stained, identified and counted. The results are reported as spores per sample or spores per gram of material. If the sample is heavily loaded with fungal material, the analyst may elect to analyze a lesser percentage of the sample preparation and extrapolate the result for the entire sample. Therefore, a sample with a significantly elevated fungal count may be an approximation.

SECTION 3 TERMS AND CONDITIONS:

3.1 Sample Retention:

Safety Environmental Laboratories and Consulting, Inc. will retain all samples for a period of 90 days. The Client may reclaim the samples during this time period. Following this time period, all samples will be disposed of in an appropriate manner.

3.2 Health Related Issues:

Safety Environmental Laboratories and Consulting, Inc. makes no written or verbal claims or recommendations as to direct health related issues based on this data or report. Clients should consult with a licensed and board certified health care professional / physician such as an allergist, immunologist, or environmental health specialist.

3.3 Insurance Claims:

Safety Environmental Laboratories and Consulting, Inc. makes no claims or recommendations as to the Client's insurance coverage. This data or report does not imply coverage of these issues by the insurance carrier.

3.4 Independent Contractor:

Safety Environmental Laboratories and Consulting, Inc. is an independent contractor and is not an employee of the Client. The Client is hereby contracting with Safety Environmental Laboratories and Consulting, Inc. to perform analytical services. Safety Environmental Laboratories and Consulting, Inc. reserves the right to determine the method, manner and means by which the services will be performed. The order or sequence of the work shall be under the control of Safety Environmental Laboratories and Consulting, Inc.

3.5 Limited Warranty:

Safety Environmental Laboratories and Consulting, Inc. warrants to Client that the material, analysis, data, programs and services, will be of the kind and quality designated and will be performed by qualified personnel. Special requirements for format or standards to be followed shall be attached as an exhibit and must be executed by both parties. Safety Environmental Laboratories and Consulting, Inc. makes no other warranties, whether written, oral or implied, including without limitation, warranty of fitness for purpose or merchantability.

This report is based on data / information supplied by the Client. Safety Environmental Laboratories and Consulting, Inc. assumes no responsibility for errors or omissions on the Client's behalf.

In no event, shall Safety Environmental Laboratories and Consulting, Inc., its employees, ownership, managers or directors be liable for special or consequential damages, either in contract or tort, whether or not the possibility of such damages have been disclosed to Safety Environmental Laboratories and Consulting, Inc. or could have been reasonably foreseen by Safety Environmental Laboratories and Consulting, Inc.

These results only apply to samples tested with client provided information. Please see attached chain of custody.

SECTION 4 REFERENCES AND SUPPLEMENTAL INFORMATION:

1. Bioaerosols: Assessment and Control, Janet Macher, Ed., American Conference of Governmental Industrial Hygienist, Cincinnati, OH 1999.
2. The Inside Story, A Guide to Indoor Air Quality, United States Environmental Protection Agency and the United States Consumer Product Safety Commission, Washington, DC (1995)
3. Exposure Guidelines for Residential Indoor Air Quality, Environmental Health Directorate, Health Protection Branch, Health Canada, Ottawa, Ontario (1989)
4. Fungal Contamination in Public Buildings: Health Effects and Investigative Methods, Health Canada, Ottawa, Ontario (2004)
5. S500 Standard and Reference Guide for Professional Water Damage Restoration, 3rd Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA (2006)
6. S520 Standard and Reference Guide for Professional Mold Remediation, 1st Edition, Institute of Inspection, Cleaning, and Restoration Certification, Vancouver, WA (2004)
7. Field Guide for the Determination of Biological Contaminants in Environmental Samples, American Industrial Hygiene Association, Fairfax, VA (2005)

8. Standards of Practice for the Assessment of Indoor Environmental Quality, Volume 1: Mold Sampling; Assessment of Mold Contamination, Indoor Environmental Standards Organization (2002)
9. "Mold Remediation: Building Assessment, Restoration, and Demolition", U.S. Occupational Health and Safety Administration (OSHA).
10. Guidelines on Assessment and Remediation of Fungi in Indoor Environments, New York City Department of Health, http://www.nyc.gov/html/doh/html/epi/moldrpt1.html.
11. "Mold Remediation in Schools and Commercial Buildings", United States Environmental Protection Agency, http://www.epa.gov/iaq/molds/mold-remediation.html.
12. Mold Resources, United States Environmental Protection Agency, http://www.epa.gov/iaq/pubs/moldresources.html.
13. "A Brief Guide to Mold, Moisture, and Your Home" United States Environmental Protection Agency, http://www.epa.gov/iaq/molds/moldguide.html.
14. "The Facts About Mold", American Industrial Hygiene Association (AIHA), http://www.aiha.org/content/accessinfo/consumer/factsaboutmold.htm
15. Worker Protection Information, United States Occupational Safety and Health Administration (OSHA), http://www.osha.gov/sltc/molds/index.hmtl
16. CDC Mold Facts, http://www.cdc.gov/mold/faqs.html

SECTION 5 ANALYTICAL RESULTS AND CHAIN OF CUSTODY:

The results of the analyses performed are attached and presented in table format. Each sample type (spore traps - air samples, surface wipes - sterile swabs, tape lifts, bulk dusts and bulk building materials are provided on separate tables. A copy of the submitted chain of custody form is provided following the analytical results.

Safety Environmental Laboratories and Consulting, Inc. appreciates the opportunity to provide these services. If you have any questions concerning this report, feel free to contact us at (205) 823-6200.

Sincerely,

Christy McKee

Christy McKee
Laboratory Director

Brad Stiles
Director of Operations

Safety Environmental Laboratories and Consulting, Inc.

Fungal Air Sample Analysis Report

989 Yeager Pkwy.
Pelham, AL 35124
Phone: (205) 823-6200
Fax: (205) 823-9066

SELC

Customer: **SELC**
989 Yeager Parkway
Pelham, AL 35124
SELC Project #: **2025-3659**

Analysis Date: **12/30/25**
Report Date: **12/30/25**
Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **136 N. Court St.**
Prattville, AL 36067
PO Number: **None Given**

Lab ID # 100766

Sample Number	**1**				**2**				**3**			
Sample Identification	6 Pod				2 Pod				3 Pod			
Sample Volume (liters)	75				75				75			
Debris Rating	3				3				4			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria	1	40	40	<1	1	40	40	<1				
Arthrinium												
Ascospores	23	920	40	9	8	320	40	4	17	680	40	7
Aspergillus/Penicillium - like	5	200	40	2	36	1,440	40	17	56	2,240	40	23
Basidiospores	146	5,840	40	56	100	4,000	40	47	109	4,360	40	45
Bipolaris/Dreschlera												
Chaetomium									1	40	40	<1
Cladosporium	81	3,240	40	31	68	2,720	40	32	55	2,200	40	23
Curvularia												
Epicoccum									1	40	40	<1
Fusarium												
Nigrospora	1	40	40	<1								
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*	3	120	40	1	2	80	40	1	1	40	40	<1
Stachybotrys												
Torula												
Trichoderma												
Ulocladium												
Other Spores	1	40	40	<1								
Hyphal Fragments/ cm^2	15	600	40		10	400	40		18	720	40	
Total Fungal Spores/ m^3		10,440				8,600				9,600		

Annie Chang

Analyst
Annie Chang - Assistant Laboratory Director

Katie Brown

Technical Review
Katie Brown - Laboratory Techinician

Safety Environmental Laboratories and Consulting, Inc.

Fungal Air Sample Analysis Report

989 Yeager Pkwy.
Pelham, AL 35124
Phone: (205) 823-6200
Fax: (205) 823-9066

SELC

Customer: **SELC**
989 Yeager Parkway
Pelham, AL 35124
SELC Project #: **2025-3659**

Analysis Date: **12/30/25**
Report Date: **12/30/25**
Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **136 N. Court St.**
Prattville, AL 36067
PO Number: **None Given**

Lab ID # 100766

Sample Number	**4**				**5**				**6**			
Sample Identification	7 Pod				Kitchen				Drug Detection Room			
Sample Volume (liters)	75				75				75			
Debris Rating	2				2				3			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria	2	80	40	1								
Arthrinium												
Ascospores	13	520	40	4	5	200	40	5	8	320	40	3
Aspergillus/Penicillium - like	5	200	40	2	6	240	40	6	13	520	40	5
Basidiospores	193	7,720	40	65	57	2,280	40	53	106	4,240	40	40
Bipolaris/Dreschlera												
Chaetomium									2	80	40	1
Cladosporium	86	3,440	40	29	38	1,520	40	36	134	5,360	40	50
Curvularia												
Epicoccum												
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*					1	40	40	1				
Stachybotrys									5	200	40	2
Torula												
Trichoderma												
Ulocladium												
Other Spores												
Hyphal Fragments/ cm^2	13	520	40		5	200	40		13	520	40	
Total Fungal Spores/ m^3		11,960				4,280				10,720		

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Fungal Air Sample Analysis Report

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SELC

Customer: **SELC**
989 Yeager Parkway
Pelham, AL 35124
SELC Project #: **2025-3659**

Analysis Date: **12/30/25**
Report Date: **12/30/25**
Method: ASTM D7391-20

Project Name: **Autauga County Jail**
Project Location: **136 N. Court St.**
Prattville, AL 36067
PO Number: **None Given**

Accredited Laboratory Lab ID # 100766

Sample Number	**7**				**8**				**9**			
Sample Identification	Room N113				Room 109A				Outside Ambient Air			
Sample Volume (liters)	75				75				75			
Debris Rating	2				1				1			
Fungal Spore Identification	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**	**raw ct.**	**spores/M^3**	**DL**	**%**
Alternaria									2	80	40	1
Arthrinium												
Ascospores	3	120	40	3	1	40	40	2	28	1,120	40	7
Aspergillus/Penicillium - like	10	400	40	11	1	40	40	2	25	1,000	40	6
Basidiospores	69	2,760	40	74	49	1,960	40	92	255	10,200	40	66
Bipolaris/Dreschlera												
Chaetomium												
Cladosporium	10	400	40	11	2	80	40	4	72	2,880	40	19
Curvularia												
Epicoccum									1	40	40	<1
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*	1	40	40	1					5	200	40	1
Stachybotrys												
Torula												
Trichoderma												
Ulocladium									1	40	40	<1
Other Spores												
Hyphal Fragments/ cm^2	1	40	40		3	120	40		10	400	40	
Total Fungal Spores/ m^3		3,720				2,120				15,560		

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Fungal Swab Sample Analysis Report

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Fax: (205) 823-9066

SELC

Customer: **SELC**
989 Yeager Parkway
Pelham, AL 35124
SELC Project #: **2025-3659**

Analysis Date: **12/30/25**
Report Date: **12/30/25**

Project Name: **Autauga County Jail**
Project Location: **136 N. Court St.**
Prattville, AL 36067
PO Number: **None Given**

Lab ID # 100766

Sample Number	**S1**				**S2**				**S3**			
Sample Identification	control room wall				records shelving				freezer wall			
Sample Area (cm^2)	25				25				25			
Fungal Spore Identification	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**
Alternaria												
Arthrinium												
Ascospores	2	480	240	7								
Aspergillus/Penicillium - like	19	4,560	240	68								
Basidiospores					2	18	9	67	2	16	8	100
Bipolaris/Dreschlera												
Chaetomium					1	9	9	33				
Cladosporium	6	1,440	240	21								
Curvularia	1	240	240	4								
Epicoccum												
Fusarium												
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*												
Stachybotrys												
Torula												
Trichoderma												
Ulocladium												
Other Spores												
Hyphal Fragments/ cm^2	194	46,560	240									
Total Fungal Spores/ cm^2		6,720				27				16		

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Safety Environmental Laboratories and Consulting, Inc.

Fungal Swab Sample Analysis Report

989 Yeager Pkwy.
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SELC

Customer: **SELC**
989 Yeager Parkway
Pelham, AL 35124
SELC Project #: **2025-3659**

Analysis Date: **12/30/25**
Report Date: **12/30/25**

Project Name: **Autauga County Jail**
Project Location: **136 N. Court St.**
Prattville, AL 36067
PO Number: **None Given**

Accredited Laboratory
Lab ID # 100766

Sample Number	**S4**				**S5**				**S6**			
Sample Identification	cell horizontal surfaces				C3 duct supply				fresh air intake for cells			
Sample Area (cm^2)	25				25				25			
Fungal Spore Identification	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**	**raw ct.**	**spores/cm^2**	**DL**	**%**
Alternaria												
Arthrinium												
Ascospores	1	8	8	33	2	16	8	29	2	240	120	5
Aspergillus/Penicillium - like	1	8	8	33					8	960	120	19
Basidiospores					3	24	8	43	17	2,040	120	40
Bipolaris/Dreschlera												
Chaetomium									1	120	120	2
Cladosporium					1	8	8	14	14	1,680	120	33
Curvularia												
Epicoccum												
Ganoderma	1	8	8	33	1	8	8	14				
Nigrospora												
Oidium/Peronospora												
Pithomyces												
Smuts/*Myxomycetes/Periconia*												
Stachybotrys												
Torula												
Trichoderma												
Ulocladium												
Other Spores:												
Hyphal Fragments/ cm^2	1	8	8						6	720	120	
Total Fungal Spores/ cm^2		24				56				5,040		

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Safety Environmental Laboratories and Consulting, Inc.

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Phone: (205) 823-6200
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Environmental, Health, and Safety Solutions

SELC

Mold/Fungal Chain of Custody Form

Customer: SELC Inc.
Address: 989 Yeager Pkwy
Pelham, AL 35124
Phone: Fax:
E-mail: brad.stiles@selcinc.com

Project Number: 2025 - 3659
Project Name: Autauga County Jail
Project Location: 136 N. Court St. Prattville, AL 36067
PO Number:
SELC Proj. #:

Turn-Around Time: ☐ Rush/Same Day† ☒ Next Day ☐ 2 Business Days ☐ 3 Business Days ☐ 4 Business Days ☐ Other:

† *Same day not available after 12:00 PM;*

***Results will be submitted by close of business. Business Hours: Monday-Friday 8:00am-5:00 pm (CST)*

- Field blanks should be submitted with all samples -

Sample #	Date Sampled	Sample Description (Sampling Area or Material)	Analysis Requested				Vol. (L) / Area (cm²)	SELC USE ONLY	
			Air	Swab	Bulk	Tape		#	Cond
1	12/29/25	6 Pod	☒	☐	☐	☐	75L	1	G
2	↓	2 Pod	☒	☐	☐	☐	↓	2	↓
3		3 Pod	☒	☐	☐	☐		3	
4		7 Pod	☒	☐	☐	☐		4	
5		kitchen	☒	☐	☐	☐		5	
6		Drug Detection Room	☒	☐	☐	☐		6	
7		Room N113	☒	☐	☐	☐		7	
8		Room 109A	☒	☐	☐	☐		8	
9		outside ambient air	☒	☐	☐	☐	↓	9	
51		control room wall	☐	☒	☐	☐	25cm²	10	
52		records shelving	☐	☒	☐	☐	↓	11	
53		freezer wall	☐	☒	☐	☐		12	
54		cell horizontal surfaces	☐	☒	☐	☐		13	
55		C3 duct supply	☐	☒	☐	☐		14	
56	↓	fresh air intake for cells	☐	☒	☐	☐	↓	15	↓
			☐	☐	☐	☐			
			☐	☐	☐	☐			
			☐	☐	☐	☐			

Sampled By:
Signature

Relinquished by:

Signature	Date	Time
	12/29/25	12:29 pm

Received By:

Signature	Date	Time
	12/29/25	12:40 pm

APPENDIX D

SUPPLEMENTAL FUNGAL INFORMATION

Alternaria sp. Mitosporic fungus. Hyphomycetes. Anamorphic Pleosporaceae.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 40-50 species.	Soil, dead organic debris, on food stuffs and textiles. Plant pathogen, most commonly on weakened plants.	Dry spore. Wind.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Commonly recognized. Type I allergies (hay fever, asthma). Type III hypersensitivity pneumonitis: Woodworker's lung, Apple store hypersensitivity. May cross react with Ulocladium, Stemphylium, Phoma, others.	Nasal lesions, subcutaneous lesions, nail infections; the majority of infections reported from persons with underlying disease or in those taking immunosuppressive drugs. Most species of Alternaria do not grow at 37°C.	A. alternata produces the antifungal alternariol. Other metabolites include AME (alternariol monomethylether), tenuazonic acid, and altertoxins (mutagenic).

Growth Indoors	Industrial Uses	Other Comments
On a variety of substrates. Aw=0.85-0.88 (minimum for various species)	Biocontrol of weeds and other plants.	One of the most common fungi worldwide.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows well on general fungal media. Colonies are dark olive green to brown, floccose to velvety (heavily sporulating). Colonies become pleomorphic over time, and lose the ability to sporulate with subsequent transfer.	Distinctive. Young spores or spore fragments may be confused with Ulocladium, Pithomyces, Stemphylium, or Epicoccum. (Some Alternaria species cannot be separated from Ulocladium.)	Distinctive. Readily identifiable on tape lift samples.

Ascospores

Spore category. Produced by morels, truffles, cup fungi, ergot and many micro-fungi.

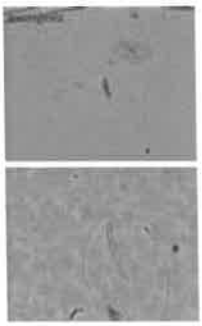
Distribution	Where Found	Mode of Dissemination
Ubiquitous. More than 3,000 genera.	Saprophytes and plant pathogens. Found everywhere in nature.	Spores are predominantly forcibly discharged during periods of high humidity or rain.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Highly variable, dependent on genus and species. Poorly studied.	Dependent on genus and species, but the vast majority do not cause disease.	Very many, dependent on genus and species.

Growth Indoors	Industrial Uses	Other Comments
The cellulolytic ascomycetes Chaetomium and Ascotricha are frequently found growing indoors on damp substrates.	Dependent on genus and species.	Some of the common asexual fungi such as Penicillium and Aspergillus produce sexual forms under certain conditions; these are classified in the ascomycete group and given distinct names. For example, the most common sexual forms of Penicillium are Talaromyces and Eupenicillium; the most common sexual forms of Aspergillus are Eurotium and Emericella.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
While some ascomycetes sporulate in culture (Chaetomium, Pleospora), many are parasitic plant pathogens, and sporulate (grow) only on living host plants.	Many ascospores are distinctive. Many others will be classified as "other colorless." In general, ascospores are recognizable by the fact that they have no attachment points, and are sometimes enclosed in gelatinous sheaths or within a sac.	Many ascomycetes are distinctive, and readily identified on tape samples, especially if fruiting bodies are present.

Photographs:



Aspergillus sp.

Mitosporic fungus. Hyphomycetes.Teleomorphs (sexual state): Eurotium, Neosartorya, Emericella (Ascomycetes).

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 200 species.	Soil, decaying plant debris, compost piles, stored grain.	Dry spore. Wind.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Common. Type I allergies (hay fever, asthma). Type III hypersensitivity pneumonitis: Humidifier lung, Malt worker's lung, Compost lung, Wood trimmer's disease, Straw hypersensitivity, Farmer's lung, Oat grain hypersensitivity, others. Other: A. fumigatus: allergic bronchopulmonary aspergillosis (ABPA), allergic fungal sinusitis.	Respiratory, invasive, cutaneous, ear, and corneal disease. Severe, invasive disease is usually associated with immunosuppressed hosts. Many species grow at 37°C (body temperature). A. fumigatus: fungus ball and invasive disease. A. flavus: nasal sinus lesions, invasive disease. A. niger: "Swimmer's ear," and invasive disease.	Partial list: A. flavus: aflatoxin B1 & B2, cyclopiazonic acid, kojic acid A. fumigatus: ergot alkaloids, fumigaclavines, gliotoxin, fumigatoxin, fumigillin, fumitremorgens, helvolic acid, tryptoquivaline tremorgens, verruculogen. A. niger: malformin C, oxalic acid. A. ustus: austocystins. A. versicolor: aspercolorin, averufin, cyclopiazonic acid, sterigmatocystin, versicolorin.

Growth Indoors	Industrial Uses	Other Comments
On a wide range of substrates. Water requirements range widely (dependent on species). Aw=0.71-0.94 (minimum for various species).	Many, including practical applications in food production. For example, A. oryzae is used to ferment soybeans to soy sauce. A. terreus produces mevinolin which is able to reduce blood cholesterol; A. niger is used in the bread and beer making industries (enzyme production) and also is able to decompose plastic. A. niger and A. ochraceus are used in cortisone production.	Aspergillus is one of the most common fungal genera, worldwide, and Aspergillus fumigatus is one of the most common species found.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Aspergillus species grow well on general fungal media. Some xerophilic species prefer dryer conditions.	Free spores are indistinguishable from Penicillium, and other genera with small round to oval colorless spores. Penicillium/Aspergillus spores may have remnants of cell wall connections.	If sporulating structures are present, Aspergillus is readily identifiable on tape samples. Old growth or samples with very large numbers of spores may not contain structures necessary for identification and are reported as "spores typical of Penicillium/Aspergillus."

Photographs:

Penicillium sp.

Mitosporic fungus. Hyphomycetes. Teleomorphs (sexual state): Eupenicillium, Talaromyces (Ascomycetes).

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 200 species.	Soil, decaying plant debris, compost piles, fruit rot. P. glabrum has been isolated from diesel fuel.	Dry spore. Wind, insects (fungus serves as a food source for storage mites).

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Common. Type I allergies (hay fever, asthma). Type III hypersensitivity pneumonitis: Cheese washer's lung, Woodman's lung, Moldy wall hypersensitivity.	One species of Penicillium species, P. marneffei, is a cause of human infection. It has not yet been found in the United States.	Various toxins by different species: penicillic acid, peptide nephrotoxin, viomellein, xanthomegin, xanthocillin X, mycophenolic acid, roquefortine C & D, citrinin, penicillin, cyclopiazonic acid, isofumigaclavine A, penitrem A, decumbin, patulin citreoviridin, griseofulvin, verruculogen, ochratoxin, chrysogine, and meleagrin.

Growth Indoors	Industrial Uses	Other Comments
Widespread. Commonly found in house dust. Grows in water damaged buildings on wallpaper, wallpaper glue, decaying fabrics, moist chipboards, and behind paint. Also found in blue rot of apples, dried foodstuffs, cheeses, fresh herbs, spices, dry cereals, nuts, onions, and oranges. Aw=0.78-0.86 (minimum for various species).	Roquefort and camembert cheese, salami-sausages starter culture; anti-bacterial antimicrobial penicillin, and anti-fungal antimicrobial griseofulvin.	Penicillium is one of the most common fungal genera, worldwide. Microbial volatile organic compounds (MVOCs) produced: Penicillium commune produces 2-methyl-isoborneol, a heavy musty odor.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows readily on general fungal media. Colonies are usually shades of blue, green, and white.	Free spores are indistinguishable from Aspergillus and other genera with small round to oval colorless or slightly pigmented spores. Penicillium/Aspergillus spores may have remnants of cell wall connections.	Penicillium is readily identifiable on tape samples if sporulating structures are present. Old growth or samples with high numbers of spores may not exhibit sporulation structures necessary for identification and are therefore reported as "spores typical of Penicillium/Aspergillus."

Basidiospores

Spore category. Produced by mushrooms, puffballs, shelf fungi, rusts, smuts, and many other fungi.

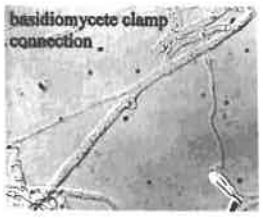
Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 1,200 genera.	Saprophytes and plant pathogens. Gardens, forests, woodlands.	Wind; spore release (active mechanism) during periods of high humidity or rain.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Probably common. Type I allergies (hay fever, asthma). Type III hypersensitivity pneumonitis: Lycoperdonosis (puffball spores), Mushroom culture hypersensitivity.	Asexual forms may cause rare opportunistic infections. The yeast Cryptococcus neoformans is a basidiomycete.	Mushroom toxicosis (poisoning) is usually a result of ingestion of the following toxins: amanitins, monomethyl-hydrazine, muscarine, ibotenic acid, psilocybin.

Growth Indoors	Industrial Uses	Other Comments
Serpula lacrimans, the agent of "dry rot," and other fungi causing white and brown wood rot, grow and destroy the structural wood of buildings. Poria incrassata causes a particularly destructive dry rot in buildings.	Many mushrooms are edible, and very important in the food industries.	Occasionally, a benign, non-wood rotting mushroom will fruit inside a building, growing in some unique ecological niche if enough moisture is present. If mushrooms are found growing indoors we ask clients to submit the entire mushroom for identification.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Most Basidiomycetes will not fruit on laboratory media. Many will form arthrospores or sterile mycelia on laboratory media.	Most basidiospores have a distinctive asymmetrical attachment point. Many basidiomycetes have recognizable spores. Serpula, the agent of dry rot, with tan-orange basidiospores, can sometimes be identified on spore trap slides.	Except for the occasional finding of Serpula (above), basidiospores are rarely found on tape lifts, except as a part of normal influx of outdoor spores.

Photographs:



Chaetomium sp.

Ascomycete.

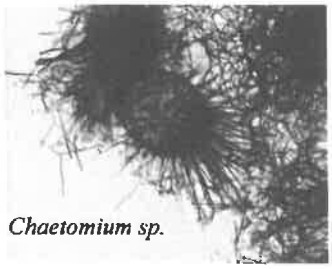
Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 81 species.	Soil, seeds, cellulose substrates, dung, woody and straw materials.	Spores are formed inside fruiting bodies. Spores are forced out an opening and spread by wind, insects, water splash.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Not well studied. Type I allergies (hay fever, asthma).	Uncommon agent of onychomycosis (nail infection).	Chaetomin. Chaetomium globosum produces chaetoglobosins. Sterigmatocystin is produced by rare species. Other compounds produced (which may not be mycotoxins in the strict sense) include a variety of mutagens.

Growth Indoors	Industrial Uses	Other Comments
Widespread, cellulolytic, very commonly found on damp sheetrock paper.	Used in textile testing and the production of cellulase.	None.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows and sporulates on general fungal media, may need 8-20 days for fruiting body production and sporulation.	Distinctive. Chaetomium globosum has small brown "lemon" or "football-shaped" ascospores.	Distinctive and readily identifiable on tape lifts.

Photographs:



Chaetomium sp.

Cladosporium sp.

Mitosporic fungus. Hyphomycetes. Teleomorphs (sexual state): Mycosphaerella, Venturia (Ascomycetes).

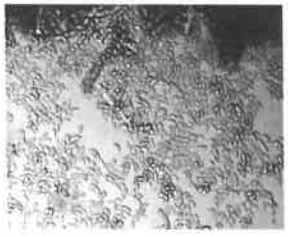
Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 28-40 species. One of the most common genera, worldwide.	Soil of many different types, plant litter, plant pathogen, leaf surfaces, old or decayed plants.	Dry spore (formed in very fragile chains, easily dispersed). Wind.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Common and important allergen. Type I allergies (hay fever, asthma). Type III hypersensitivity pneumonitis: Hot tub lung, Moldy wall hypersensitivity.	Generally, non-pathogenic. One species, Cladosporium carrionii, is an agent of chromoblastomycosis in subtropical and tropical regions (grows at 35-37°C).	Cladosporin, emodin. (Neither are highly toxic.)

Growth Indoors	Industrial Uses	Other Comments
Widespread, on many substrates, including textiles, wood, moist window sills. Grows at 0°C, and so is associated with refrigerated foods. Aw=0.85-0.88 (minimum for various species).	C. herbarum produces enzymes which are used in the transformation of steroid intermediates such as pregnenolone and progesterone, biologically important hormones used in the industrial production of oral contraceptives.	G.S. deHoog & J. Guarro have placed species associated with human infection in a new genus Cladophialophora, i.e. Cladophialophora carrionii, C. bantiana. Older medical texts refer to this fungus by its former name Hormodendron species.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows on all general fungal media. Some species sporulate better than others, and some may need cycles of light in order to produce spores.	Distinctive, with wide variation in size and shape. Spores with dark attachment scars and some olive to brown pigmentation are identified as Cladosporium.	Distinctive, readily identifiable on tape lifts.

Photographs:



Curvularia sp.

Mitosporic fungus. Hyphomycetes. Teleomorph (sexual state): Cochliobolus (Ascomycete).

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. More commonly found in tropical, subtropical regions. Approx. 30 species.	Plant debris, soil, facultative plant pathogens of tropical or subtropical plants.	Dry spore. Wind.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Common. Type I allergies (hay fever, asthma). Other: A relatively common cause of allergic fungal sinusitis.	Occasionally a cause of onychomycosis, ocular keratitis, sinusitis, mycetoma, pneumonia, endocarditis, cerebral abscess, and disseminated infection. Most cases are from immunocompromised patients.	Not known.

Growth Indoors	Industrial Uses	Other Comments
Yes, on a variety of substrates.	Not known.	None.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows well on general fungal media; most isolates need "light/dark cycling" for sporulation. Colonies are shades of gray to brown.	Distinctive; large second or center cell gives conidia pronounced curved shape. Conidia from species with less pronounced curve may be misidentified. Some Drechslera spores are similar.	Distinctive, readily identifiable on tape lifts.

Epicoccum sp.

Mitosporic fungus. Hyphomycetes.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Two species.	Plant debris, soil. Secondary invader of damaged plant tissue.	Dry spore. Wind. Spores also released by hygroscopic movement.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Common. Type I allergies (hay fever, asthma).	No cases of infection have been reported in humans or animals.	Antibiotic substances produced: flavipin, epicorazine A & B, indole-3-acetonitrile.

Growth Indoors	Industrial Uses	Other Comments
Yes, on many different substrates including paper, textiles, and insects. Aw=0.86-0.90 (minimum).	None known.	None.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows well on general fungal media, although sporulation may be strain dependent. Colonies typically have orange reverse pigment.	Intact spores are distinctive. Young spores or spore fragments may be confused with Ulocladium, Stemphylium or possibly Alternaria. Commonly found in outdoor air.	Distinctive, readily identifiable on tape lifts.

Nigrospora sp.

Mitosporic fungus. Hyphomycetes. Teleomorph (sexual state): Khuskia (ascomycete).

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Especially abundant in warm climates. Approx. 4-5 species.	Decaying plant material and soil.	Active discharge mechanism. Does not require wind or rain.
Allergen	**Potential Opportunist or Pathogen**	**Potential Toxin Production**
Type I allergies (hay fever, asthma).	Very rare report of human infection.	Not known.
Growth Indoors	**Industrial Uses**	**Other Comments**
Rarely found growing indoors.	Not known.	None.
Characteristics: Growth/Culture	**Notes on Spore Trap Recognition**	**Notes on Tape Lift Recognition**
White, floccose, spreading. Develops black spore clusters with time.	A distinctive large, dark brown (nearly black), globose spore is readily identifiable on spore trap slides.	Distinctive, but rarely found.

Smuts

Fungal category. Ustilaginales. Basidiomycetes.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Two families, 50 genera, and 950 species.	On cereal crops, grasses, weeds, other fungi, and on other flowering plants.	Wind disperses the powdery brown teliospores of smut.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Type I allergies (hay fever, asthma).	No reports of human infection by the plant parasitic forms.	Not known.

Growth Indoors	Industrial Uses	Other Comments
Smuts do not usually grow indoors. They are parasitic plant pathogens that require a living host for the completion of their life cycle.	Not known.	Smuts are members of the Basidiomycetes and have two spore types: teliospores (dry, powdery stage) and basidiospores (yeast stage).

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
The airborne phase (teliospores) of smut requires a living host for growth and will not develop on laboratory media. The yeast phase (basidiospores) is saprophytic and will grow on general fungal media.	Smut teliospores cannot easily be distinguished from the myxomycetes and certain species of Periconia. They are reported in the "round, brown" spore category: "Smuts, Periconia, myxomycetes."	The teliospores of smuts are somewhat distinctive en masse. They are found in dust as part of the normal influx of outdoor particles

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Myxomycetes

Taxonomic fungal category. Slime molds.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 45 genera.	Decaying logs, stumps and dead leaves, particularly in forested regions.	These organisms have both dry and wet spores. Wind disperses the dry fruiting body spores, whereas the wet amoebic phase is motile.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Type I allergies (hay fever, asthma). (Lycogala used in one skin test survey.)	No reports of human infection.	None.

Growth Indoors	Industrial Uses	Other Comments
Occasionally found indoors.	None known.	The myxomycetes have an interesting life cycle which includes a wet spore phase and a dry spore phase. When conditions are favorable, they move about like amoebae, resembling primitive animals. When conditons are not favorable they form a resting body (sclerotium) with dry, airborne spores. The myxomycetes are not considered to be true fungi.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
The myxomycetes do not grow on general fungal media.	While a few are distinctive, many of the myxomycete spores are difficult to distinguish from the smuts. These spores are placed in our group "smuts, myxomycetes, Periconia," due to their similar "round, brown" morphology.	Occasionally seen and identified on tape lifts. Distinctive especially when fragments of the lacy fruiting bodies are present.

Periconia sp.

Mitosporic fungus. Hyphomycetes.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 20 species.	Soil, blackened and dead herbaceous stems and leaf spots, grasses, rushes and sedges. Almost always associated with other fungi.	Dry spore. Wind.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Not studied.	Rare case of mycotic keratitis reported.	Not known.

Growth Indoors	Industrial Uses	Other Comments
Rarely found growing indoors.	Not known.	None.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Colonial morphology is similar to Cladosporium. Periconia is infrequently isolated in culture.	Some species have distinguishing spore characteristics and are recognizable. Generally it is difficult to distinguish Periconia spores from the smuts, myxomycetes and other round, brown spore types.	Spores with underlying sporulating structures are distinctive, although we very rarely see them on tape lifts.

Stachybotrys sp.

Mitosporic fungus. Hyphomycetes.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 15 species.	Soil, decaying plant substrates, decomposing cellulose (hay, straw), leaf litter, and seeds. Growth not influenced by soil pH or copper; growth enhanced by manure.	Wet spore. Insects, water splash. Wind when dried out.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Not well studied. Type I allergies reported.	No reports of human infection. (No species grow well at 37°C.)	Macrocyclic trichothecenes: verrucarin J, roridin E, satratoxin F, G & H, sporidesmin G, trichoverrol; cyclosporins, stachybotryolactone. Stachybotrys mycotoxicosis: human toxicosis has been described; may be characterized by dermatitis, cough, rhinitis, itching or burning sensation in mouth, throat, nasal passages and eyes. The best described toxicoses are from domestic animals that have eaten contaminated hay and straw or inhaled infected material from contaminated bedding.

Growth Indoors	Industrial Uses	Other Comments
Commonly found indoors on wet materials containing cellulose, such as wallboard, jute, wicker, straw baskets, and other paper materials. (See "Characteristics: Growth/Culture"). Aw=0.94	Not known.	Many human reports of Stachybotrys toxicosis are anecdotal. Stachybotrys mycotoxicosis is currently the subject of toxin research.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows well on general fungal media. Stachybotrys is slow growing as compared to Penicillium and other common mold genera, and may not compete well in the presence of other fungi. However, when water availability is high for prolonged periods on environmental material, Stachybotrys may gradually become the predominating mold, especially on cellulose containing materials.	Spores of the species S. chartarum are distinctive, and not easily confused with other genera. Carbon fragments which may be oval and of similar size may sometimes be confused with S. chartarum. Memnoniella and Gliomastix produce spores with similar gray black pigment. Note: Spore trap samples are more likely to demonstrate the presence of Stachybotrys than culturable samples (Andersen).	Distinctive, readily identifiable on tape lift samples. Direct microscopic observation of samples is often necessary as Stachybotrys may be missed if only culture methods are used.

Ulocladium sp.

Mitosporic fungus. Hyphomycetes.

Distribution	Where Found	Mode of Dissemination
Ubiquitous; cosmopolitan. Approx. 9 species.	Soil, dung, paint, grasses, fibers, wood, decaying plant material, paper, and textiles.	Dry spore. Wind.

Allergen	Potential Opportunist or Pathogen	Potential Toxin Production
Major. Type I allergies (hay fever, asthma). Ulocladium cross-reacts with Alternaria, adding to the allergenic burden of Alternaria-sensitive patients.	Rare subcutaneous tissue infection.	Not known.

Growth Indoors	Industrial Uses	Other Comments
Widespread. Found on gypsum board, paper, paint, tapestries, jute, other straw materials. Ulocladium has a high water requirement.	Not known.	None.

Characteristics: Growth/Culture	Notes on Spore Trap Recognition	Notes on Tape Lift Recognition
Grows well on all general fungal media. Colonies are dark brown to rusty brown, granular to velvety. Geniculate sporulating structures can be observed with the stereoscope.	Distinctive brown spores. Young spores or spore fragments may be confused with Alternaria, Pithomyces, and others, although Alternaria usually has shades of olive green pigment.	Distinctive, readily identifiable on direct observation. Certain species may form rudimentary beaks and short chains which may be confused with Alternaria.

#3

IN THE CIRCUIT COURT OF AUTAUGA COUNTY, ALABAMA

State of Alabama, ex rel., MARK HARRELL Sheriff of Autauga County Alabama,	)	
Plaintiff,	)	
v.	)	
	)	**Civil Action Number:**
AUTAUGA COUNTY COMMISSION; JAY THOMPSON, Chairman, RUSHTON "RUSTY" JACKSLAND, Commissioner, JOHN THRAILKILL, Commissioner, BILL TATUM, Commissioner; and TERRY TANNER, Commissioner,	)	**04-CV-2025-900111**
Defendants.	)	

FIRST AMENDED VERIFIED COMPLAINT

COMES NOW the Plaintiff, State of Alabama, ex rel. Mark Harrell, in his Official Capacity as the Duly Appointed Sheriff of Autauga County, Alabama by and through his undersigned counsel of record, Hon. Dwight ("Tray") M. Richardson III, and Hon. William P. Gray, and files this Complaint against the Autauga County Commission as a party Defendant and each of the named Commissioners as a Party Defendant alleging a Petition for Writ of Mandamus; Declaratory Judgment; Mandatory Injunction; and for Other Related Relief against Defendants, Autauga County Commission, and Defendants Jay Thompson, Chairman; Rushton "Rusty" Jacksland, Commissioner, John Thrailkill, Commissioner, Bill Tatum, Commissioner, and Terry Tanner, Commissioner, All in their Official Capacity, and would show the following unto this Honorable Court:

THE PARTIES:

1. The Plaintiff, State of Alabama, ex rel, Sherriff Mark Harrell, is the duly appointed to fill the unexpired term of his predecessor Sheriff of Autauga County and is represented by Hon. Dwight ("Tray") M. Richardson III, ASB#6247, G68R, Phone: (334) 414-4321 Email: tray@dwightrichardson.com and William P. Gray, ASB#-5268-R78W, Phone (205) 968 – 0900 Email: wpg@grayattorneys.com.

2. The Defendants, Autauga County Commission, which acts though its Chairman, Hon. Jay Thompson, in his Official Capacity.

3. The Defendant, Rushton "Rusty" Jacksland, who is an Autauga County Commissioner, in his Official Capacity.

4. The Defendant, John Thrailkill, who is an Autauga County Commissioner, in his Official Capacity.

5. The Defendant, Bill Tatum, who is an Autauga County Commissioner, in his Official Capacity.

6. The Defendant, Terry Tanner, who is an Autauga County Commissioner, in his Official Capacity.

VENUE AND JURISDICTION:

7. This Court is the proper venue and has jurisdiction of the parties and the subject matter herein.

INTRODUCTION:

8. This case involves the negligent or willful refusal of the Defendant Autauga County Commission and the Defendant Commissioners to follow their statutory mandated duties to maintain the Autauga County Metro Jail. Further at the heart of this suit is the Autauga County Commission's apparent willful refusal to follow their statutory mandated duty to provide the Sheriff of Autauga County, Alabama, with all necessary funds to fulfill his statutory mandated duties.

9. The Petitioner, Sheriff Mark Harrell, has tried since his appointment as Sheriff of Autauga County, by Gov. Kay Ivey on January 16, 2023, along with his predecessors (Sheriff Herbie Johnson) going back as far as 2006, to provide Autauga County, Alabama, a safe and suitable jail and to maintain said jail in a safe and suitable condition. Alabama Code § 11-14-10 mandates that "The County Commission shall erect courthouses, jails, and hospitals and other necessary county buildings, and such county commission shall have authority to levy a special tax for that purpose. Each county within the state shall be *required to maintain* a jail within their county." Additionally, Alabama Code § 36-22-18 states the following: "the County commission shall also furnish the Sheriff with the necessary quarters, books, stationary, office supplies, postage and other conveniences and equipment, including automobiles and necessary repairs, maintenance and all expenses incidental thereto, as are reasonably needed for the proper and efficient conduct of the affairs of the Sheriff's office."

10. Alabama law mandates that the **Sheriff** has authority over the jail, and this authority is independent of the County Commission pursuant to Alabama Code §14-6-1. Alabama law also provides that the duties of the Counties with respect to jails are *limited* to providing all necessary funds for the proper and efficient conduct of the affairs of the Sheriff's Office, the operation of the

jail, and to provide the necessary facilities to house the jail and maintain the jail. Ala. Code § 36-22-18. The County Commission cannot substitute its judgment for that of the Sheriff.

11. In this matter, however, the Autauga County Commission has repeatedly failed to grant the request of the sheriff to hire necessary staff, APOST certified deputies, and other personnel and further has failed to provide the sheriff with the necessary staff, contractual employment, or other maintenance personnel to properly clean and maintain the jail, and to provide and maintain courthouse security and courtroom security.

12. As far back as September 2006 then Lieut. Larry Nixon, pursuant to the budget request for the [Sheriff] at that time, requested funds to hire a maintenance person in order to keep and maintain the Autauga County Metro Jail (ACMJ) properly and to clean the jail from mold and mildew which had been recognized as a problem almost every time the health department had visited the jail. Under Budget Line items 216 and 219, then Lt. Nixon requested particularly under line item 216 the money to combat health department warnings about mold and mildew.

13. The request for maintenance personnel, or, alternatively, a maintenance contract has been repeated several times and has never been granted. All of these matters took place long before Sheriff Mark Harrell was appointed on or about January 16, 2023, as Sheriff of Autauga County by Gov. Kay Ivey. Since that time Plaintiff has repeatedly tried in good faith to provide a safe jail for the citizens of Autauga County, Alabama. Additionally, the Commission has repeatedly failed to communicate on a regular basis with Sheriff Mark Harrell.

THE FACTUAL BACKGROUND:

14. The Plaintiff, Mark Harrell, is the Sheriff of Autauga County. Pursuant to article I, Section 14 of the Constitution of Alabama 1901, he is a member of the Executive Branch of the State of

Alabama. He is the Chief Executive Officer of the Autauga County Sheriff's Office (ACSO). *Hale v. Randolph Co. Comm* 423 So. 2d 893 (Ala. Civ. App. 1982). Plaintiff is conversant with the facts set out herein and the same are true and correct to the best of the Plaintiff's knowledge and belief.

15. Defendants are the Autauga County Commission and the individual Commissioners, Jay Thompson, Chairman, Rushton "Rusty' Jacksland, Commissioner, John Thrailkill, Commissioner, Bill Tatum, Commissioner; and Terry Tanner, Commissioner (hereinafter, Commission, Defendants). They are all subject to the provisions of Alabama Code § 11-1-1, et seq. (1975) and further subject to Alabama Code § 36-22-18, et seq. (1975). Suit is being brought against Defendants, the Autauga County Commission, and Defendants Thompson, Jacksland, Thrailkill, Tatum, and Tanner, in their official capacities.

16. Sheriff Harrell, and his predecessors in office have tried for years to eliminate and replace the dangerous mold and mildew found in the Autauga County Metro Jail. There have been repeated attempts by Sheriff Harrell, and his predecessors, to persuade the commission Defendants to fulfill their mandated, required, statutory, and constitutional duty to properly fund the needed maintenance of the Autauga County Metro Jail (ACMJ). These requests go as far back as February 2018 with the discovery of condensation on the air supply grills for the HVAC system in the Autauga County Metro Jail. Finally, the most recent requests were made on April 17, 2025 and April 25, 2025 (attached hereto as Exhibits "A" and "B"). The predecessor to Sheriff Harrell received a report from Honeywell who, while in the process of doing an inspection, discovered the "potential" for mold and in February 2019 Environmental Materials Consultants, INC out of Montgomery, Alabama, at the request of the Autauga County Commission, at that time, did a complete report of mold testing at the Autauga County Metro Jail and found a high concentration of mold of all different kinds and also provided certain remediation requirements.

17. In February 2023 the County Administrator, Scott Kramer, received a detailed email of proposed jail changes from Capt. Larry Nixon concerning the ACMJ and in March of the same year the Autauga County Risk Services Report outlining problems in the Autauga Metro Jail was received. In August 2023 a second EMC Mold Report was received which did not contain any remediation. Subsequently, Capt. Nixon also obtained a cleaning quote from ProTek, LLC, for mold cleaning at the Autauga Metro Jail. That quote was forwarded to the Defendant County Commissioners and in September 2023 the Sheriff and Capt. Nixon made the first request to the County Commission that this entire matter be put on the agenda for an executive session to discuss all of the jail problems and the issues involved therein. However, to the dismay of the Plaintiff, no executive session was granted, nor was there any general discussion regarding the problems that had been identified and determined to exist with regard to mold in the Autauga County Metro Jail.

18. From April through August 2023 other events had occurred regarding the results of the inspection of the sprinkler system and letters were exchanged between Dr. David McMichael and Audra Smith in reference to the findings that had been made in inspections of the Autauga County Metro Jail and the problems resulting therefrom. No action was taken at that time.

19. On March 26, 2024, Capt. Nixon sent emails from the Sheriff's Office and included responses to the emails, as well as the invoice that had been received when "black" mold was discovered in the Autauga County Metro Jail. On May 3, 2024, Capt. Nixon detailed and advised the Sheriff, Mark Harrell, of the course of events that had taken place in April of 2024. On the same day, after receipt of the email from Capt. Nixon, Sheriff Harrell sent emails to Scott Kramer and all County Commissioners regarding the discovery of mold in the air vents in the Autauga County Metro Jail. At that time Sheriff Harrell requested for the second time, an executive session for emergency funding and remediation. However, once again to the dismay of Sheriff Harrell and

Capt. Nixon, Defendant Commissioners failed to grant any such request. On May 6, 2024, Capt. Nixon received emails from Scott Kramer requesting that Capt. Nixon contact several named companies for mold remediation. On May 7, 2024, Sheriff Harrell sent an email to Scott Kramer regarding what quotes had been obtained.

20. On May 8, 2024, Capt. Larry Nixon sent an email to Scott Kramer with regard to mold remediation and received an email reply from Scott Kramer asking for the thoughts of Patrick Addison (PH and J) for the price of mold remediation. On May 14, 2024, almost two months after the confirmation of the discovery of "black" mold having been discovered in the Autauga County Metro Jail, the Autauga County Health Department sent a letter stating that, on or about May 9, 2024, a *complaint* had been received by the Autauga County Health Department concerning these matters. Thereafter, on May 10, 2024, an inspection was completed, and mold was indeed found in the Autauga County Metro Jail, as well as, other concerns.

21. On May 14, 2024, Servpro sent a letter regarding work that had been completed in cleaning the vents in the HVAC system and pictures of what they had found, as well as an invoice for the cleaning. Between May 23, 2024 and May 25, 2024 Scott Kramer sent emails that had been exchanged regarding ARK Remediation, but ARK Remediation failed to show up at the jail on the day designated. On May 25, 2024, Capt. Nixon sent an email to the County Commission and Scott Kramer that ARK Remediation had not arrived as previously designated. On May 30, 2024, Sheriff Harrell sent another email to the County administrator, Scott Kramer, requesting any information on the jail air system and information on when such remediation and work would be able to start. Sheriff Harrell received no response.

22. On May 31, 2024, Hon. Dwight ("Tray") M. Richardson III, attorney for Sheriff Harrell, contacted County attorney Kyle Shirley and informed him of the situation and that the existence

of the mold in the Autauga County Metro Jail was very significant, and that Sheriff Harrell may have no other option other than to close the jail and evacuate all prisoners. In that communication Hon. Dwight ("Tray") M. Richardson III requested once again that an executive session be held regarding all of these matters. Once again, no response was ever received.

23. EMC sent an email from Regina Mims for an open purchase order to evaluate the ductwork at the Autauga County Metro Jail and on May 31, 2024, a purchase order for EMC for evaluation of the ductwork was issued in the amount of five hundred dollars ($500.00).

THE CLOSING OF THE JAIL:

24. On June 4, 2024, the state fire marshal conducted an inspection of the jail at the request of Sheriff Harrell. At the County Commission meeting conducted on the evening of June 4, 2024, attorney Josh Pendergrass was informed that the Sheriff might have no other choice but to close the Autauga County Metro Jail due to the existence of, and the extent of, the mold that had been discovered in the jail. Additionally, subsequent to that meeting the state fire marshal issued a report that was given to Sheriff Harrell that the Metro Jail be placed on "fire watch" due to the issues that had been discovered, including the mold that was contained in the Autauga County Metro Jail. Additionally, the state fire marshal reported that there were corroded sprinkler heads and there was no functioning fire alarm at the time. On June 6, 2024, Sheriff Harrell ordered the Autauga County Metro Jail evacuated for health and safety reasons that had existed but had not been addressed. All inmates that were at that time in the Autauga County Metro Jail were transported to other safe locations in Blount County, Cullman County, Lowndes County and Russell Counties with a cost to house them of two hundred twenty-two thousand five hundred eighty-five dollars and seventy-eight cents ($222,585.78).

25. On June 10, 2024, a purchase order was obtained by Audra Smith in reference to the duct cleaning by CW Smith Decorating for five thousand dollars ($5000). On the same day, June 10, 2024, EMC completed their report on the Autauga County Metro Jail which showed mold throughout the jail, and which also contained information regarding proper remediation. At the request of Scott Kramer, Capt. Nixon escorted the EMC contingent through the ACMJ and they tested eight random samples in the Autauga County Metro Jail. The sheriff was informed that the samples were shipped overnight to EMSL Analytical, INC, in Smyrna, Georgia which is an accredited laboratory, who was requested to perform a microscopic examination to help identify the various mold spore types that were present in the Autauga County Metro Jail. Cladosporium mold was found in all eight sample locations in "medium to high" spore count, Aspergillus/Penicillium were reported at three of the locations with Penicillium\Talaryces at one location. Stachybotrys Memnoniella ("Black" Mold) was found at one location.

26. Due to the actions of Sheriff Mark Harrell in evacuating the Autauga County Metro Jail it is believed that health hazards to jail personnel, deputies, and staff, as well as health hazards to inmates and others visiting the Autauga County Metro Jail, have been minimized. The actions taken by Sheriff Harrell were done to prevent injuries to deputies, staff, and potentially others including inmates. The costs of remediation for the present mold was determined to be three hundred seventy-eight thousand dollars ($378,000.00) by A+ Cleaning and Restoration INC. That did not include the replacement of all ductworks. All requests from the Sheriff's Office from either Sheriff Harrell or Capt. Nixon were made pursuant to Ala. Code § 36-22-18, as well as other statutory code sections including Ala. Code § 11-12-15 for the benefit of the citizens of Autauga County and others who may be either incarcerated in the Autauga County Metro Jail or work as a part of the staff, deputies, or be a visitor to the ACMJ pursuant to Ala. Code § 36-22-18.

27. Subsequently, Sheriff Harrell sought to have A+ supervise the initial remediation after the very first workers showed up without personal protective equipment and were observed attempting to clean the mattresses. All fabric, including the mattresses, fabric furniture, etc. must be properly disposed of. The presence of the Sheriff and Kevin Ricke of A+, who requested the remediation protocols, resulted in a letter from the county attorney and Defendants threatening criminal action against Mr. Ricke and A+ for "interfering with the work." No one was "interfering" with any work. Just trying to ensure all work was completed correctly. Moreover, no protocols were ever provided by Defendants, and it appears that the remediation was not done according to known protocols.

28. Ala. Code § 36-22-18 has mandated that the County commission must fund these aspects of the Sheriff's office: stating as follows: "The county commission shall also furnish the sheriff with the necessary quarters, books, stationery, office supplies, postage and other conveniences and equipment, including automobiles and necessary repairs, maintenance and all expenses incidental thereto, as are reasonably needed for the proper and efficient conduct of the affairs of the Sheriff's Office." The commission Defendants publicized requests for bids and entered into a contract for remediation of the "first phase". That contract was in excess of one hundred fifty thousand dollars ($150,000.00) above the estimate that A+ had submitted. The commission Defendants have reported that everything has been completed satisfactorily for the first phase, however, despite the request of Sheriff Harrell for testing of all the areas that had previously been tested and any further known areas to be tested using the same lab that EMC had used for tests in Smyrna, Georgia. The Defendant commission has failed to do so. Mold, is a living organism and unless properly tested a mere "walk-through" and viewing of the previous mold areas is not satisfactory.

DEPUTIES TO PROVIDE COURTHOUSE SECURITY:

29. In April 2025 Hon. Judge Joy Pace Booth, Circuit Judge for the 19th Judicial Circuit in Autauga County, Chilton County, and Elmore County requested courthouse security in and around the Autauga County Courthouse, as well as, courtroom security in the courthouse. In June 2023 Judge Booth had requested and obtained an inspection and report from the Marshall of the Alabama Appellate Courts, Alabama Supreme Court, Earl Marsh, detailing his assessment of the courthouse security in and around the courthouse facility. Courthouse security and courtroom security is required inside and outside of the Autauga County Courthouse. Additionally, during an inspection conducted by the Sheriff's Office in April of 2025 it was learned that there is *no fire alarm facility* inside the Autauga County courthouse. These matters must be remedied. While commission Defendant Thrailkill and Defendant Thompson attended the meeting with Judge Booth, the Autauga County Commission has declined to grant Sheriff Harrell's requests for additional APOST certified deputies, salary, and benefits, and the necessary vehicles to replace the high mileage vehicles in the Sheriff's Office necessary to provide the requested security without reducing some of the Sheriff's Office services.

30. The Sheriff's Office for Autauga County, Alabama is taking steps to immediately develop and begin the requested court room and courthouse security requested by Judge Booth, by providing overtime to APOST certified deputies to provide the requested courthouse and courtroom security, but unless the requested additional 20 APOST certified deputies and the necessary funding for those deputies and the necessary vehicles, the Sheriff's Office will have no alternative but to reduce some of the services which are necessary to provide the reasonably necessary proper and efficient affairs of the Sheriff's Office. Courthouse and courtroom security being sensitive topics which must be provided, Plaintiff has requested on numerous occasions to

meet in executive session to discuss and provide for these matters, but all requests have thus far been denied by Defendant Autauga County Commission and the Defendant Commissioners as stated herein. Additionally, Sheriff Harrell has expended more than one hundred twenty thousand dollars ($120,000.00) since June 2024 in attempting to get the proper remediation and testing and removal of the mold infestation, provide for courthouse and courtroom security, to increase the pay for APOST certified deputies in Autauga County to meet equally the pay of surrounding counties and to increase the number of deputies necessary to provide for the safety welfare and law enforcement personnel in Autauga County, Alabama as are reasonably needed for the proper and efficient conduct of the affairs of the Sheriff's Office. Sheriff Harrell respectfully requests that upon a final hearing in this Hon. Court, that this Hon. Court order Defendants to repay the aforesaid sum as well as, all other costs, fees, and expenses expended by the Sheriff in this matter.

31. Autauga County Sheriff's deputies are being paid less than police officers in the Millbrook Police Department, the Prattville Police Department, the Montgomery Police Department, the Wetumpka Police Department, the Clanton Police Department, and the Selma police Departments. Autauga County is a large County and has a population in excess of 60,000 persons which is more people than Butler County, Chilton County, Dallas County, and Jackson County.

32. There are other staffing needs that are required by the Sheriff's Office for the proper and efficient conduct of the affairs of the Sheriff's Office which must be funded as set forth in Ala. Code § 11-8-3 and 11-12-15 all for the benefit of and the safety, and security of the citizens of Autauga County. The County Commission has failed to provide for these needs. Sheriff Harrell is informed and believes that other expenditures were budgeted and made that are lower in priority,

pursuant to Ala. Code § 11-12-15 including payment of the Defendant Commission salaries. As of the time of the filing of this complaint the commission Defendants have failed to complete the remediation of ACMJ facilities and have failed to address the need for an increased number of qualified deputies and jailers. The needed proper pay for the deputies, and an increase in the number of deputies that are necessary to provide for the safety, welfare, and law enforcement personnel in Autauga County, Alabama are reasonably needed for the proper and efficient conduct of the affairs of the Sheriff's Office. All requests and notices have been made pursuant to Ala. Code § 36-22-18 for the safety, welfare, and benefits to the citizens of Autauga County and the failures of the commission Defendants are failing to provide for those benefits and protection for the citizens of Autauga County. All of the duties set forth in Ala. Code § 36-22-18 are mandatory and required duties of the commission Defendants and their continued disregard thereof are both dangerous, and in disregard of their constitutionally mandated duties and a failure to fulfill their oath of office.

THE PRIORITY BY STATUTE:

33. By law, the Defendants must budget essential services in order of their statutory priority. The Alabama Legislature has determined the priority of essential services, and priority of funding by the Defendants as follows:

§11-12-15. Preferred claims and order of their priority; payment of same.

(a) the following claims are declared to be preferred claims against the county, and they shall be given priority in the order named:

(1) costs of heating the county jail or supplying it with wholesome water for drinking and bathing, of keeping it in a cleanly condition and free from invasive odors and of providing it with

necessary water closets and dry earth, beds, bedding and clothing; fuel; water; light; janitor's services of the courthouse and jail;...."

(2) Compensation of the members of the county commission; compensation of deputy sheriffs, the probate judge, the sheriff, the tax assessor, the county treasurer and jailers for services performed by them and authorized to be paid by them by law;...."

34. In view of the priority of the statutory mandated requirement of Ala. Code §11-12-15, the Autauga County Metro Jail and the maintenance and safety provisions are to be provided before the payment of compensation of the Autauga County Commission.

35. There are other matters contained in the remainder of Ala. Code §11-12-15, which are material and relevant and set forth therein, including the needs of the Sheriff's office to have funds to maintain and keep clean and safe the Autauga County Metro Jail for the benefit and protection of the citizens of Autauga County. This is a statutory mandate. It cannot legally be avoided. The Defendant County commission and Defendant Commissioners also must maintain communication with the Sheriff concerning what is necessarily required for the county jail because the Defendant County Commission is without authority to substitute its own judgment about what is necessary to properly run and maintain the County jail. As far back as 1987 the Atty. Gen. of the state of Alabama issued an opinion to the sheriff of Franklin County, Alabama regarding the following question:

"Does the Franklin County commission have the authority to tell the sheriff what he needs and the types of products he needs and or to refuse a purchase requisition?" The Atty. Gen. responded in part as follows:

"The County commissioners cannot substitute their judgment for that of the Sheriff regarding what is needed for the efficient operation of the Sheriff's Office.

36. Sheriff Harrell, on behalf of the state of Alabama and the citizens of Autauga County is informed and believes that sufficient funds and revenue exists at the present time to forthwith complete the remediation of the Autauga County Metro Jail, including the removal of all of the HVAC ductwork and replacement and complete the proper correction and mediation without delay and to provide, as well, the additional deputies, automobiles, jailers and other items requested to provide courthouse security and courtroom security, and other items that are reasonably needed for the proper and efficient conduct of the affairs of the Sheriff's Office. This must include proper testing of all areas of the Metro jail. The Defendant County commission has repeatedly failed to address the jail requests submitted by the Sheriff and Capt. Nixon and have not given any plausible reason for such failure. The Defendant Commission has continued to substitute its judgment for that of the Sheriff regarding the safety and welfare of the citizens of Autauga County and the proper and efficient conduct of the affairs of the Sheriff's Office in Autauga County, Alabama and thus have failed to abide by Ala. Code § 11-8-3 and 36-22-18 as well as 11-12-15. Thus, the named County Defendants are in violation of the duties imposed upon them under the Alabama code sections set forth herein. This failure to perform the duties required of the Defendant County commission is a failure to perform a ministerial duty and to perform a clear legal duty imposed by law and is detrimental to the operation of the Sheriff's Office and to the citizens of Autauga County, Alabama.

COUNT ONE: WRIT OF MANDAMUS

37. The Plaintiff incorporates all of the above paragraphs by this reference.

38. The Plaintiff State of Alabama ex rel., Sheriff Mark Harrell, prays that this Honorable Court expedite a hearing on the Jail maintenance and proper staffing thereof and hold a trial if necessary. At the conclusion of which, this Court is requested to issue a proper Writ of Mandamus ordering the Defendant Commissioners and Commission Defendants to follow all legally applicable statutory mandates herein and provide for the necessary maintenance and funding either by contract, or hiring additional personnel, to properly clean, maintain, and remediate the Autauga County Metro Jail including all the HVAC systems and the HVAC ductwork and the necessary testing thereof to ensure that the work was done properly and to provide proper staffing for said jail and the Sheriff's Office. These are dire circumstances that exist. The Commission Defendants must fund the proper staffing for the Sheriff's Office and the funds to provide the continuing maintenance of the Autauga County Metro Jail, as well as, the Courthouse and Courtroom security and the needed fire alarm for the Autauga County Courthouse.

39. The Plaintiff has a clear legal right to the order sought; there is a mandatory ministerial imperative duty upon the Commission Defendants to comply with their statutory duties as supported by the facts set forth herein; and the Plaintiff is left with no other adequate remedy as Alabama law mandates that the Commission Defendants provide the funding necessary to properly remediate and maintain the Autauga County Metro Jail in a safe, clean, and proper condition and provide the requested Courthouse security and all funding necessary for the proper and efficient conduct of the affairs of the Autauga Co. Sheriff's Office. Further, Alabama law mandates that the Autauga County Commission Defendants provide the adequate staffing as requested by the Sheriff and cannot substitute their judgment for the staffing requested by Sheriff Harrell. The Plaintiff, therefor, respectfully request that this Honorable Court have a trial on this matter and grant the Writ of Mandamus ordering the Commission Defendants to fulfill their mandated statutory duties

and all such other relief in equity and good conscience as this Hon. Court shall determine including required proper testing of the Autauga Co. Metro Jail, and proper staffing to maintain said Jail and Sheriff's Office and Courthouse/Courtroom security including both Jailers and Deputies.

COUNT TWO – DECLARATORY JUDGMENT:

40. The Plaintiff incorporates all of the above paragraphs by this reference.

41. Pursuant to Ala. Code § 6-6-223, the Plaintiff respectfully requests this Honorable Court to enter a Declaratory Judgment ordering the Commission Defendants to comply with all applicable statutes and legislative mandates, including not substituting their judgment for that of the Sheriff, Mark Harrell, and including proper cleaning, testing, and maintenance, as well as proper staffing for the Autauga County Metro Jail immediately. The Plaintiff further respectfully requests that the Court declare the Order as a final judgment pursuant to Alabama Code of 1975 §6-6-222 and requests such further, other and different relief as the court in its judgment and in equity and in good conscience shall grant. The Plaintiff further requests that upon an expedited hearing the court determine all costs herein pursuant to Ala. Code of 1975 §6-6-231 including all costs expenses and attorney's fees expended by the Plaintiff, Sheriff Harrell in this matter and tax all such cost against the Defendant Commission and Defendant Commissioners in their official capacity.

42. In light of the years of delay and apparent willful disregard of their statutory duties the Plaintiff further requests that each of the commission Defendants be fined such sum as the court shall determine per month until the completion of the matters involved with the maintenance and staffing are commenced and that such fines be paid from the commission Defendant's official bonds due to their apparent willful failure and refusal to follow their mandatory statutory duties.

The Plaintiff prays for such other further and general relief to which in equity and good conscience this Hon. Court shall declare and that upon a final hearing this Hon. Court will enter an order which enforces all statutes at issue which are applicable to commission Defendants.

RELIEF REQUESTED:

WHEREFORE, Plaintiff requests an expedited hearing and demands judgment against the commission Defendants to immediately and properly fund the cleaning, remediation, and maintenance, including the hiring either by individuals or by contract of personnel sufficient to maintain the jail in a safe and proper manner, and to provide adequate staffing as determined by the Sheriff for the Courthouse and Courtroom security, the protection and welfare of the citizens of Autauga County Alabama, and to provide such funding in accordance with the determination of the budgetary needs and expenses set forth in the facts and legal priorities herein above that are reasonably needed for the proper and efficient conduct of the Autauga County Metro Jail and affairs of the Sheriff's Office, or SHOW CAUSE, IF ANY THERE BE, WHY SUCH RELIEF SHOULD NOT BE GRANTED. Further, the sheriff requests the following relief also:

(A) This matter be set down in an expedited fashion.

(B) Upon hearing this matter, issue an order allowing the Plaintiff to have an expert of his choosing perform whatever testing deemed prudent and necessary to determine whether remediation has been done properly without undue legal threat.

(C) Upon hearing this matter, the Court require the Commission Defendants fund the requested and necessary testing and fund continuing maintenance of the Autauga County Metro Jail to provide adequate staffing and continue to take the proper remedial action, including post remediation testing to confirm that remediation was done adequately and properly, and to prevent further mold infestation.

(D) Upon final hearing in this matter, order the Defendants to adequately fund the Sheriff's Office to provide for the requested courthouse security, as well as funding and making all changes set out and recommended by Marshall Marsh in his August report to Judge Booth.

(E) Upon final hearing in this matter, to order the Defendants to adequately provide for the Sheriff's Office, including the matters specified in Ala. Code § 36-22-18 including the required vehicles and other matters necessary for the proper and efficient conduct of the affairs of the Sheriff's Office.

(F) Upon the final hearing of this matter, the Plaintiff be afforded such other, further, and different relief as may be just and equitable, as this Hon. Court shall determine.

(G) Require the Defendant commission be taxed with all costs, expert witness fees, and expenses of this proceeding and order the Defendants to properly fund the future expected maintenance costs, security costs, and all expert witness fees, expenses, and attorney's fees associated with this claim and cause of action and such further other further, and different relief, as this court in equity and good conscience may deem appropriate under the circumstances.

Respectfully submitted, this ___ day of July, 2025.

VERIFICATION

STATE OF ALABAMA)
COUNTY OF Autauga)

Before the undersigned officer duly authorized to administer oaths came **Sheriff Mark Harrell**, after first being duly sworn, states that he is authorized for the purpose of verifying the facts and allegations contained in the foregoing Amended Complaint.

Dated this the 17th day of July, 2025.

Mark B. Harrell, Sheriff

Mark Harrell, Sheriff of Autauga County

Subscribed and sworn before me
this 17 day of July, 2025.

(seal)

Notary Public

My commission expires: 3-30-2028

/s/ William P. Gray, Jr.
WILLIAM P. GRAY, JR. (ASB-5268-R78W)
Attorney for Plaintiff, Mark Harrell, Sheriff of Autauga County, Alabama

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/s/ Dwight ("Tray") M. Richardson III Richardson
DWIGHT ("TRAY") M. RICHARDSON III (ASB-6247-G68R)
Attorney for Plaintiff, Mark Harrell, Sheriff of Autauga County, Alabama

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(334) 325-0010
tray@dwightrichrdson.com

CERTIFICATE OF SERVICE

I hereby certify that on the __ day of July 2025, I electronically filed the foregoing with the Clerk of Court using the Alafile electronic document filing system which sends notification of such filing to the following attorneys of record and/or placing same in the United States Mail, postage prepaid:

Kyle Shirley
McDowell, Faulk, & Shirley, LLC.
County Attorney
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kyle@mcdowellfaulk.com

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/s/ William P. Gray, Jr.

#4

AUTAUGA COUNTY SHERIFF'S OFFICE

MARK B. HARRELL, SHERIFF

162 WEST 4TH STREET; PRATTVILLE, ALABAMA 36067

PHONE: 334-361-2500 / FAX: 334-361-2515

PUBLIC SAFETY PRIORITY BUDGET REQUESTS

1. **Pay Raise** – For **ALL** Sheriff Office Employees (Sworn Deputies, Jail employees and Office Staff). The Sheriff's Office is in need of a significant pay raise of at least 15% to be competitive with other agencies within our region. The pay raise for all employees allows the Autauga County Sheriff's Office to hire highly qualified staff, jailers and Deputies. This will allow the Sheriff's Office to retain individuals that are the best available. In a competitive environment of hiring Law Enforcement and staff we need to be aggressive in trying to achieve a pay scale that no other agency is able to do to show we are serious about the Safety and Security of Autauga County. Pay should constantly adjusted to remain competitive as other agencies increase their pay. There should be incentives also for hiring Deputies already post certified and with experience as most agencies do across the state. I'm requesting local legislation to adjust the pay for the Autauga County Sheriff to be above any non-elected official of Autauga County. Any elected official of Autauga County's pay should be more than any non-elected employee and if the non-elected officials pay is increased so should the Elected Officials. Any legislation should not alter or omit any requirements for the Office of Sheriff of Autauga County.
2. **Man Power**- We need to hire at a minimum of 20 new Deputies. In light of recent events the Autauga County Sheriff's Office needs to make sure we have more Deputies to answer calls for service, Secure the Autauga County Court House and also the Autauga County Probate Office. All of which is a statutory duty of the Sheriff's Office and statutory duty of the Autauga County Commission.
3. **Vehicles-** We are in need of a minimum of 20 new Patrol vehicles. The Autauga County Sheriff's Office, at the time of this memo have 18 vehicles with over 120,000 miles. Having high mileage vehicles puts the Deputies safety at risk while responding to calls for service and also high maintenance cost for these vehicles.
4. **Training Facility-** The time is now for a Training Area/ Facility for the Autauga County Sheriff's Office. I request that the property owned by the citizens of Autauga County on County Road 41 be utilized for the Autauga County Sheriff's Office to operate a Firearms /Training Facility. This would be multi-phase project for the Sheriff's Office at our direction and would need the Engineering Department to assist in the Clearing of the Property. This is an ideal location as to being in a safe location and also to reduce the noise in the area. We have done several checks of the area and land owners in that location themselves are constantly target practicing. A training area is needed to properly train the Autauga County Deputies consistently without trying to coordinate with outside agencies to fulfill our training needs as a Law Enforcement Agency.
5. **Equipment-** Purchase of Departmental firearms that are up to date. We need to update the Firearms of the Department to modern firearms utilized by agencies throughout the United States. I request the Purchase of the Sig Sauer P320 Pro. We need at-least 50 pistols and Rifles. This is to upgrade to a dependable and reliable weapon system for the safety of our Deputies.
   * 10 in-car printers
   *10 Radars
   *20 new radios for patrol vehicles
   * $60,000 for Office upgrades and improvements for the security of the Sheriff's Office

These are all vital requests to not only make sure the Deputies of Autauga County are able do their jobs safely and efficiently, but provide the best service available for the citizens of Autauga County. We are one of the most underfunded agencies within our state by population and growth.

Mark B. Harrell

Sheriff Mark B. Harrell